

# Phytoplankton Physiology Seminar

*Week 1, Session 0*

*Mick Follows, Instructor*

*Arianna Krinos, Teaching Assistant*



## Some logistics!

- Meet 1 time/week for 1.5 hours (Fridays 1-2:30pm)
- Other than participation in weekly discussions, students are required to lead the discussion approximately twice over the course of the semester
- 1-2 papers will be discussed per week, and each will have a discussion leader

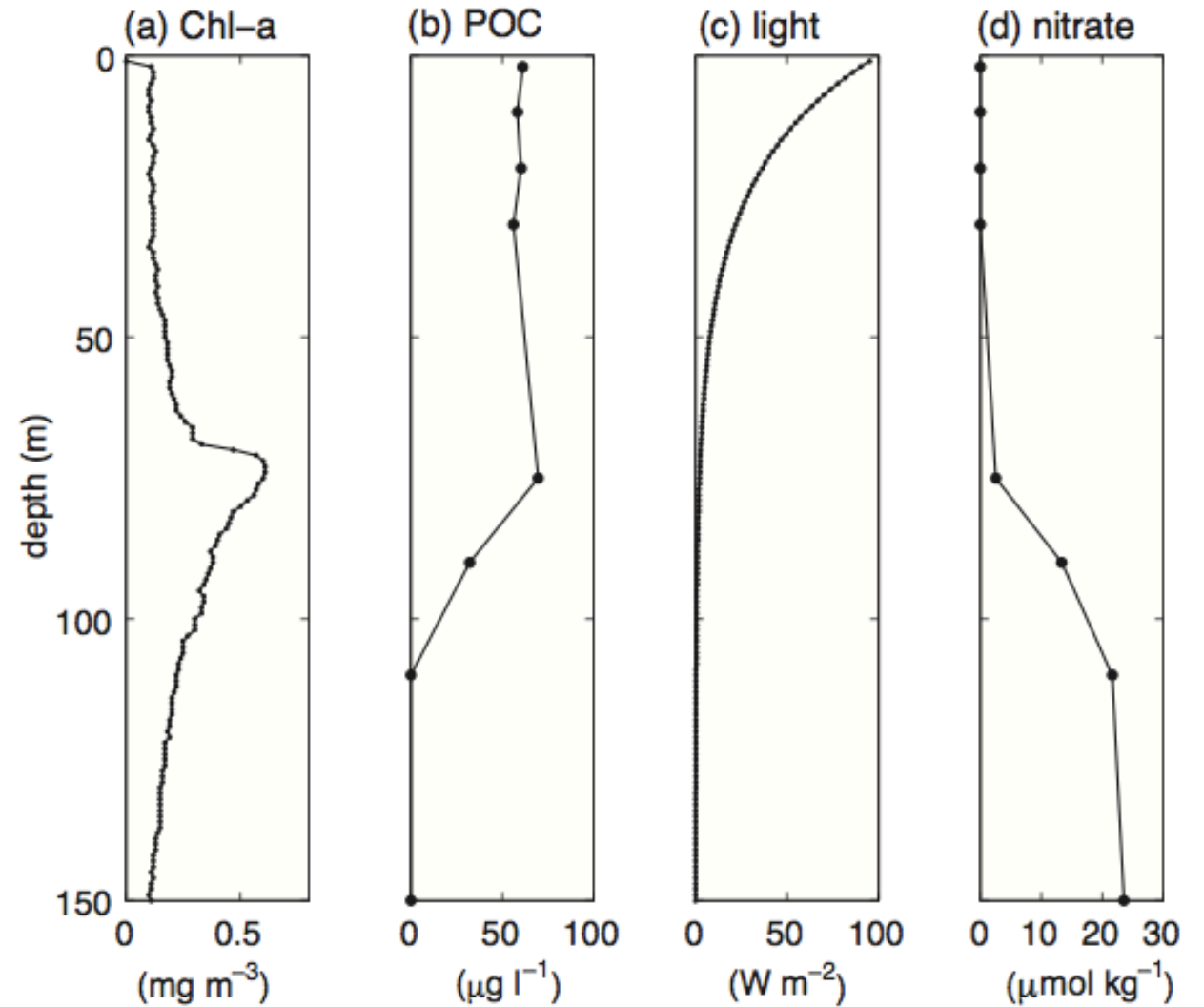
# Landscape of this brief introduction

- I. The ocean context
- II. The microbial loop + biological carbon pump
  - I. Why phytoplankton, though small, are mighty in global ocean ecology + even physics and chemistry
- III. A primer on phytoplankton sizes, functions, and physiological features
- IV. An introduction to major taxonomic groups

# The oceanic context for phytoplankton communities

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Light and mixing in the upper ocean enables phytoplankton growth



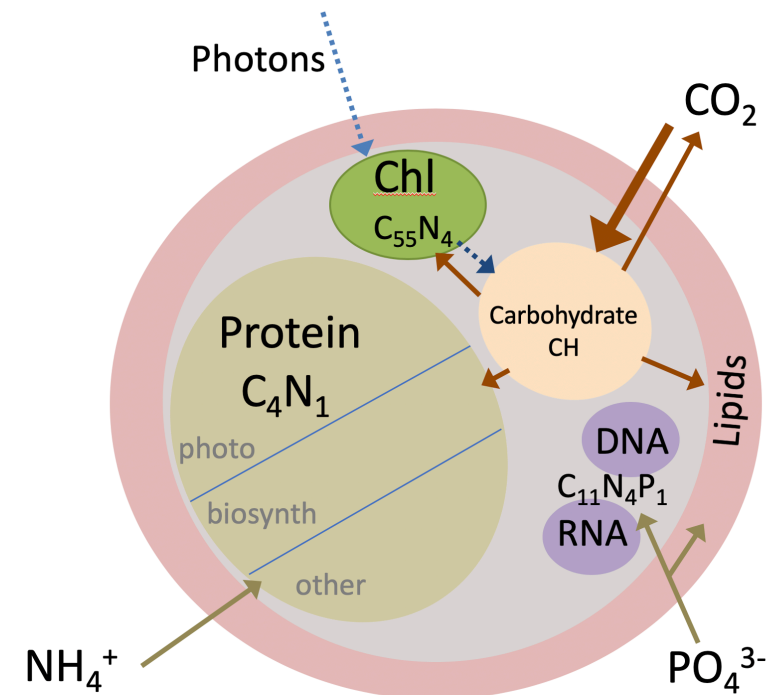
*Average* molar elemental  
composition of phytoplankton

The Redfield Ratio  
describes average  
particle nutrient ratios

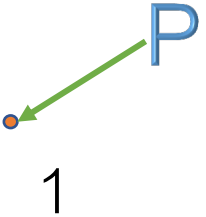
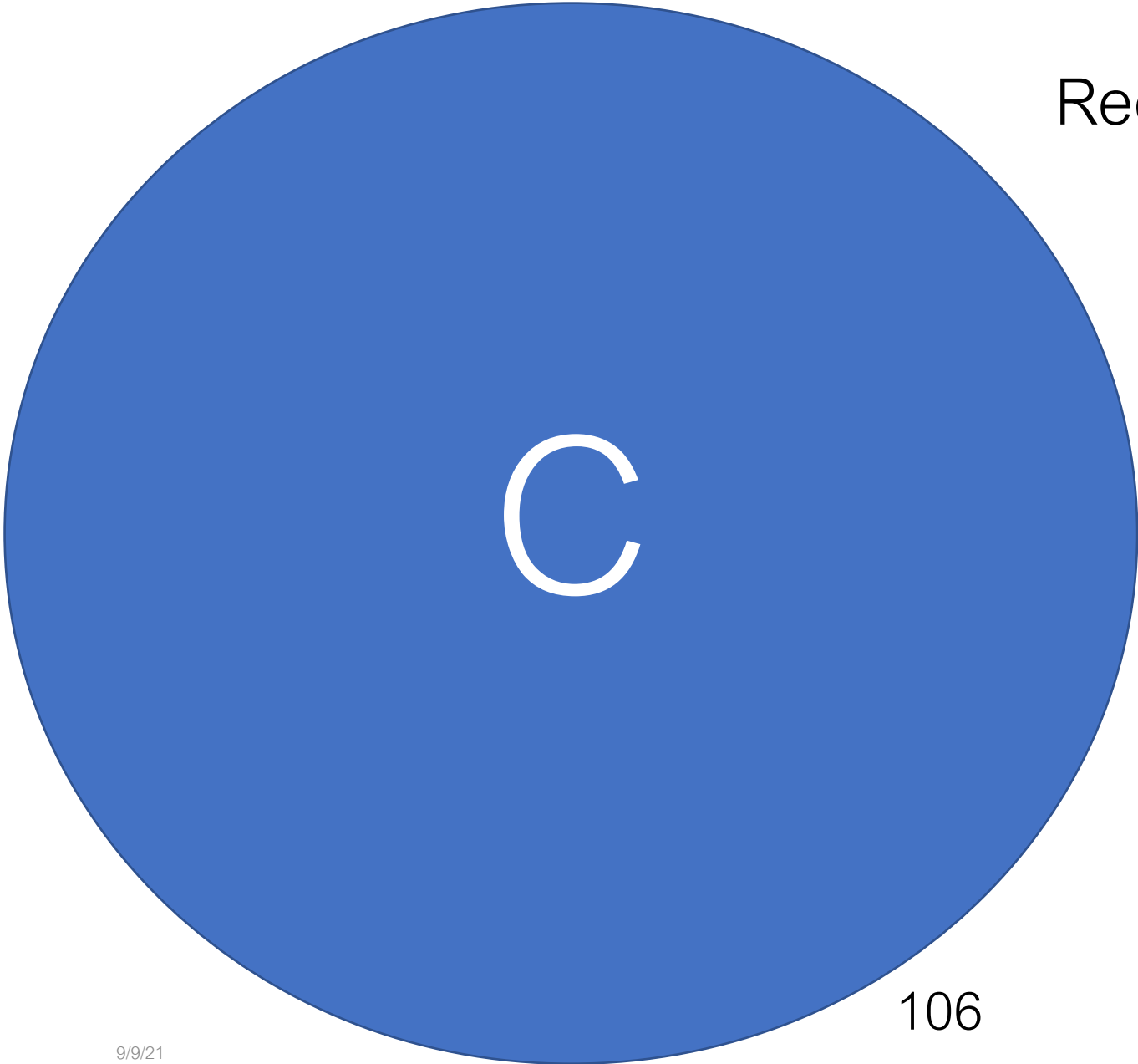


Notes:

- Highlighted by Alfred Redfield in 1930s
- $\text{O}_2$  estimated by Redfield from Redox/thermodynamic considerations
- Recent authors find the values aren't always constant

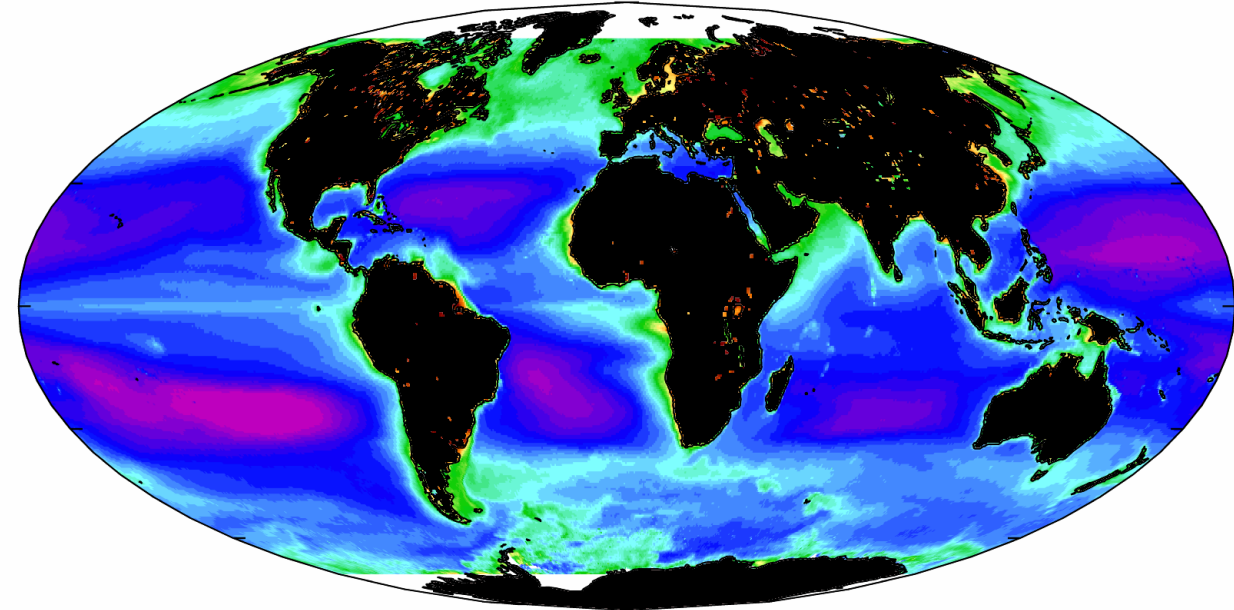
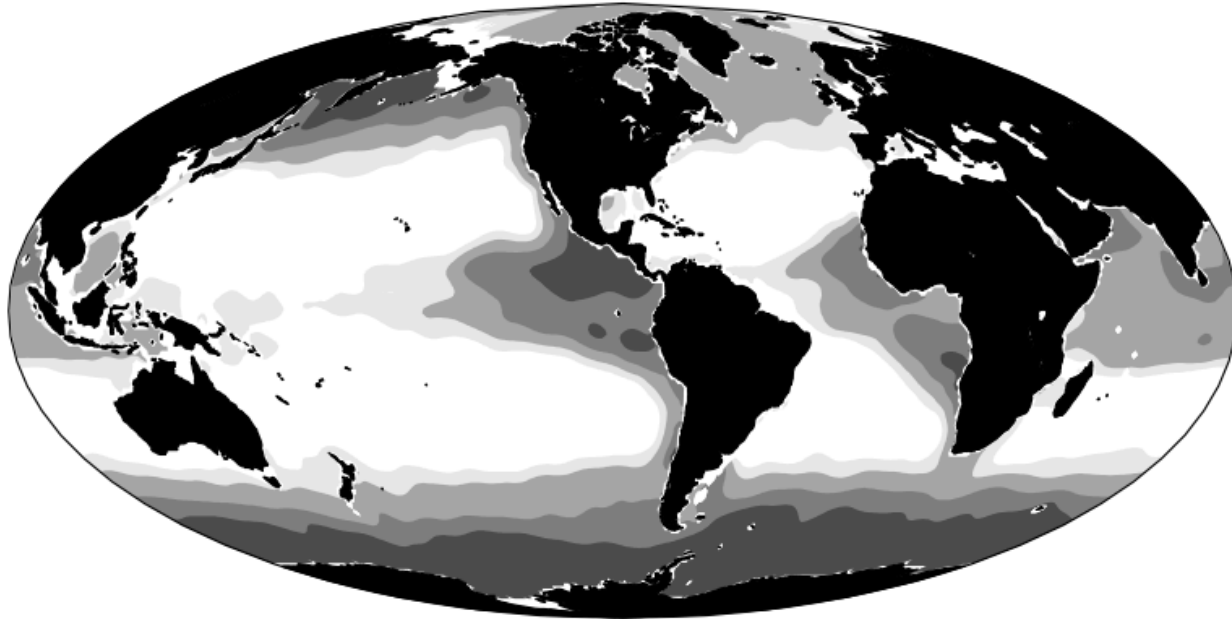


# Redfield Ratio

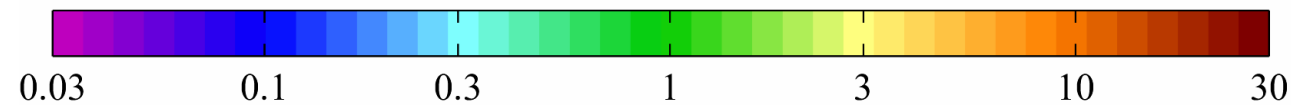


Near surface nitrate concentration reflects pattern of upwelling

nitrate at 100m ( $\mu\text{M}$ )



Average sea-surface chlorophyll, 1998 to 2006 [ $\text{mg chl m}^{-3}$ ]



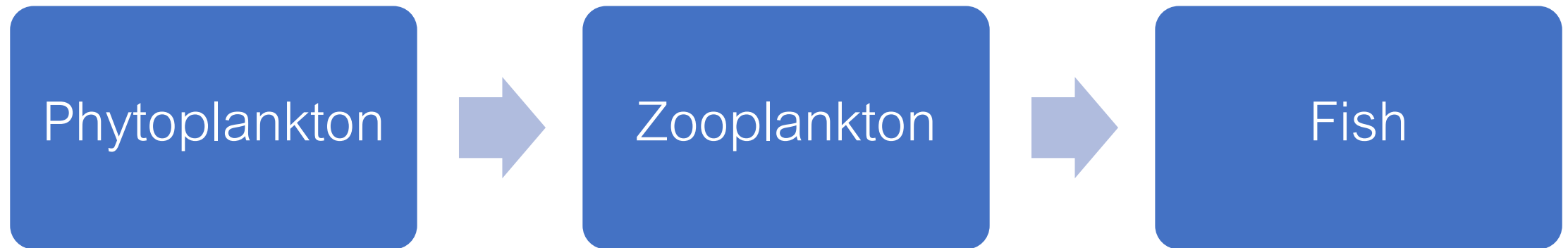
Carbon doesn't typically limit phytoplankton growth – other nutrients do



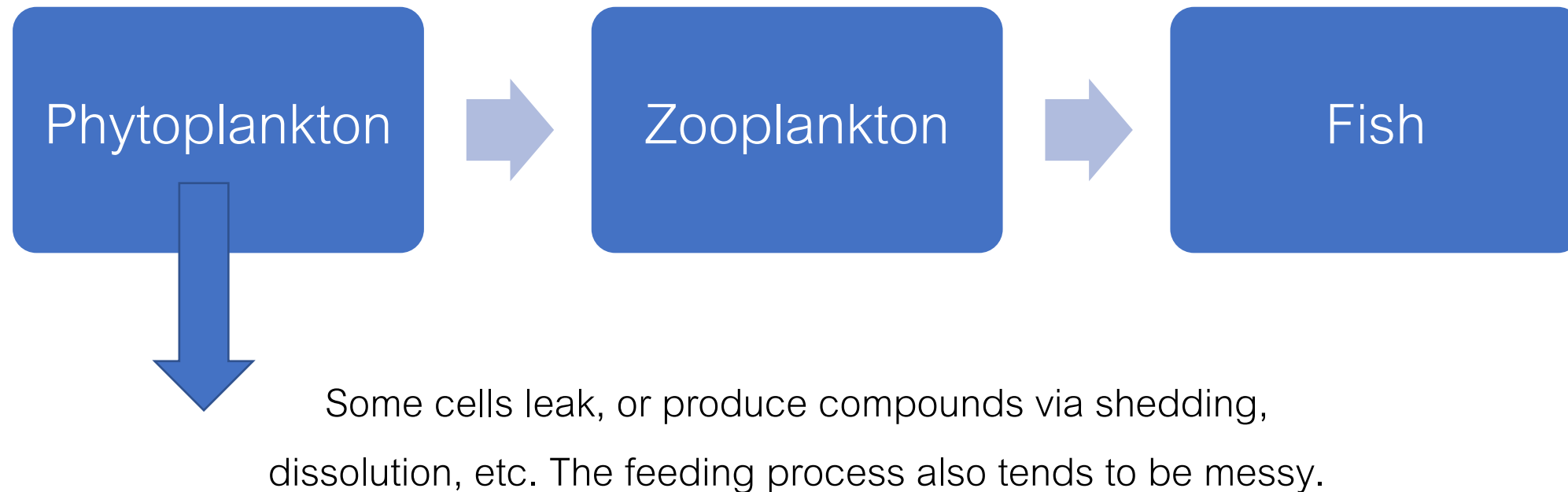
# Why are phytoplankton important?

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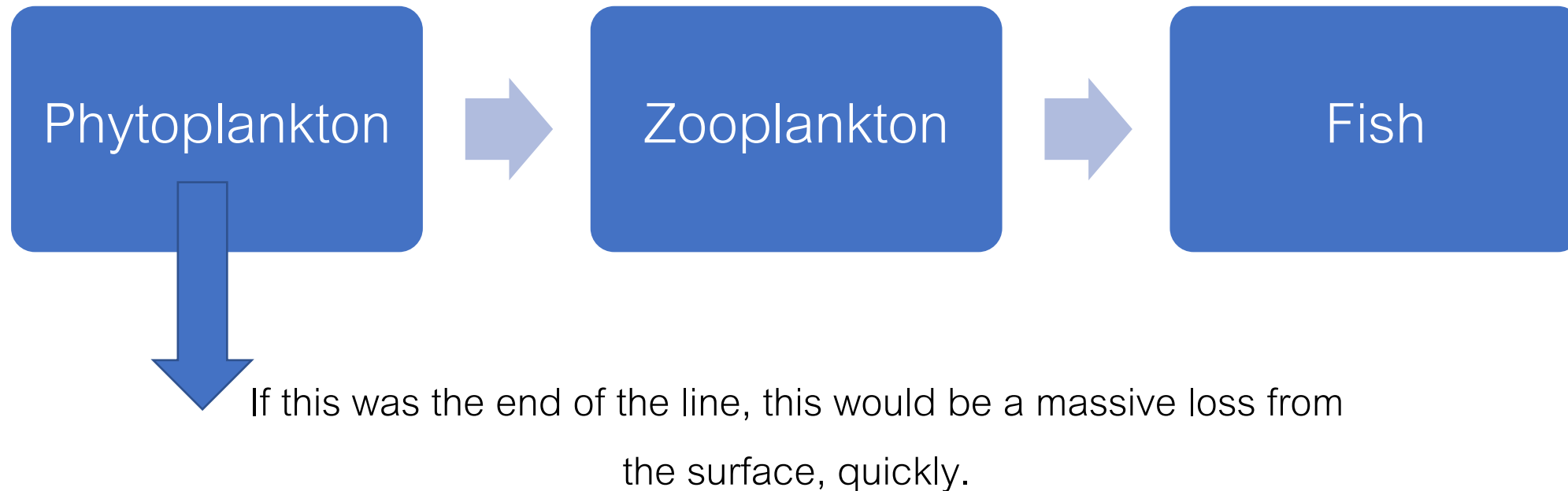
# The traditional paradigm of the ocean food web



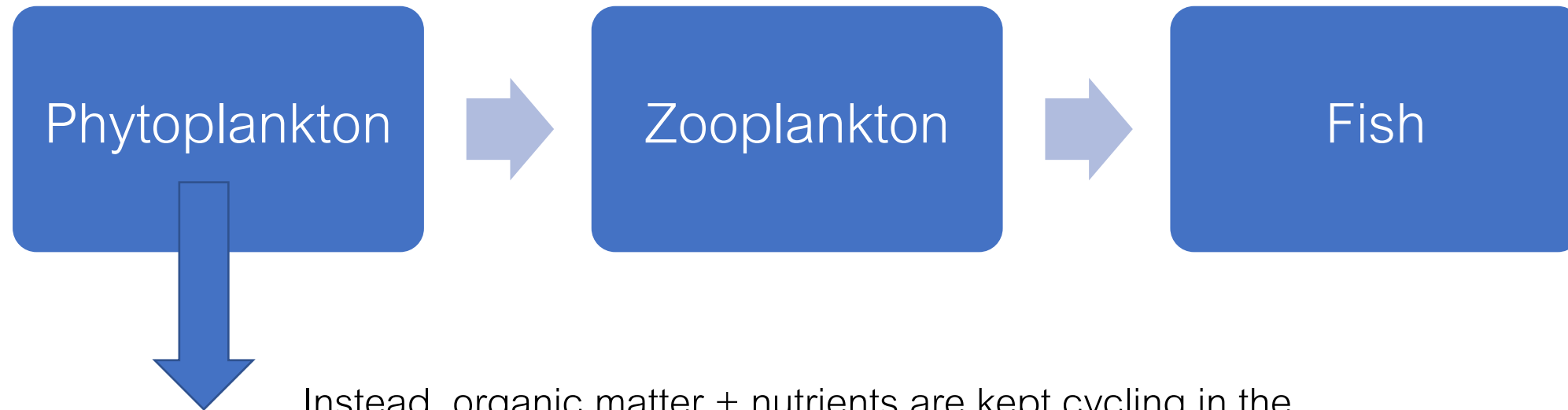
It turns out that this oversimplifies things...



It turns out that this oversimplifies things...

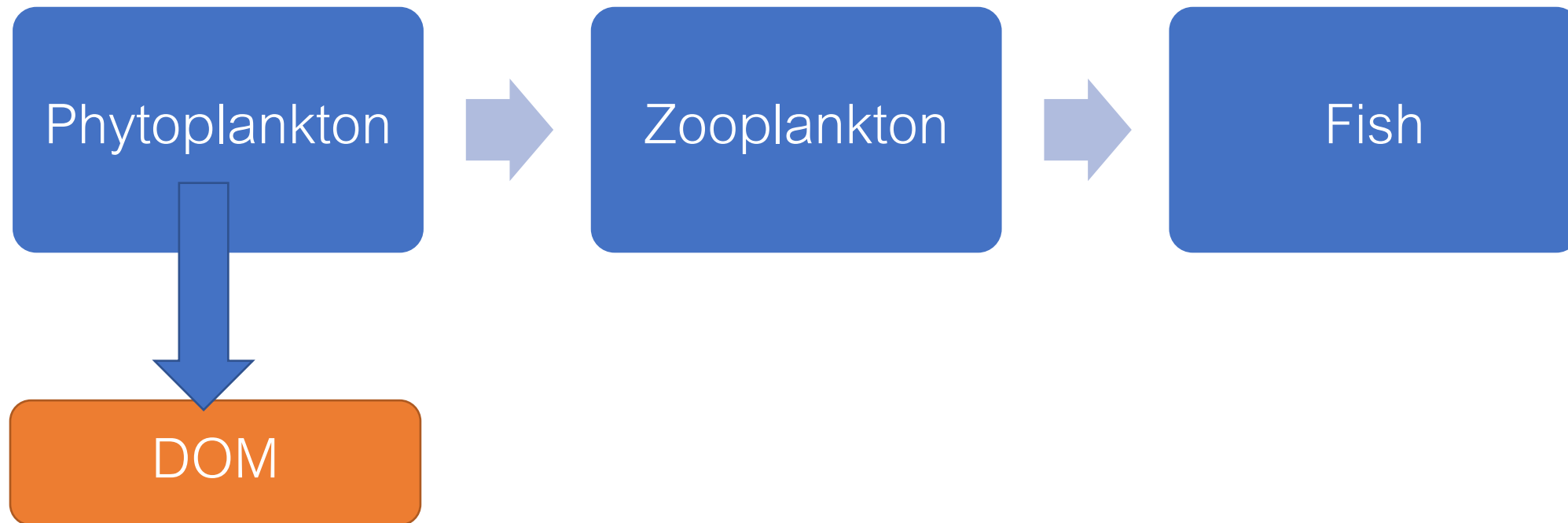


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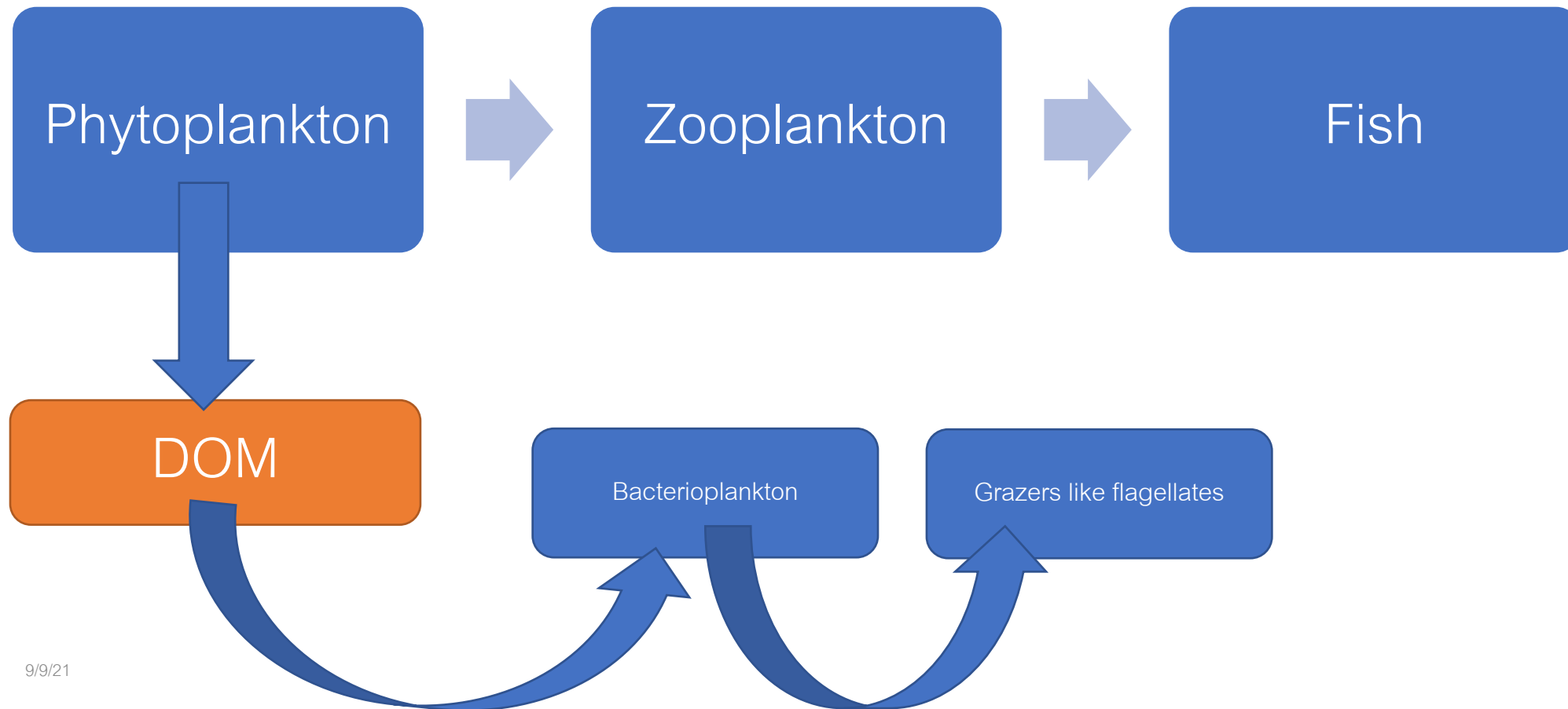


Instead, organic matter + nutrients are kept cycling in the productive surface oceans via the microbial loop.

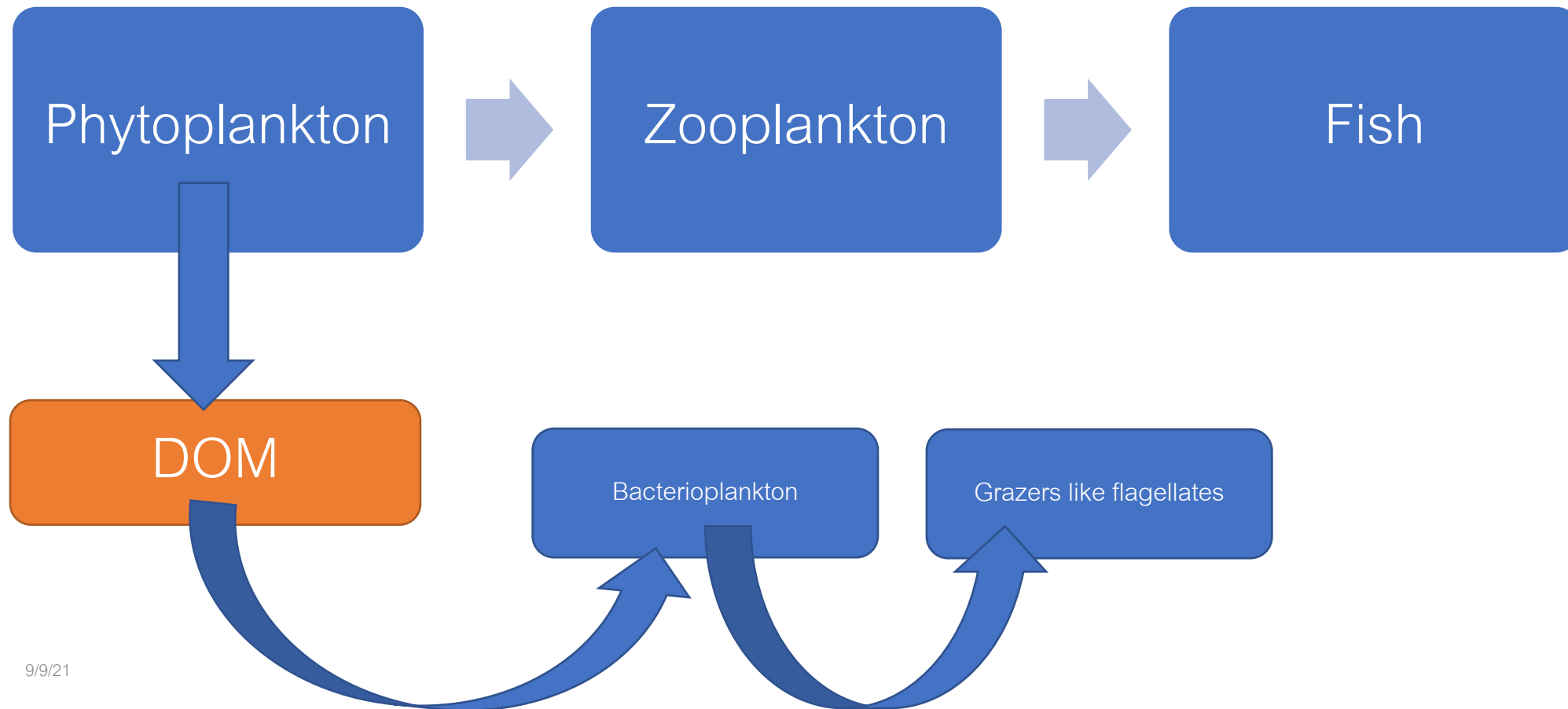
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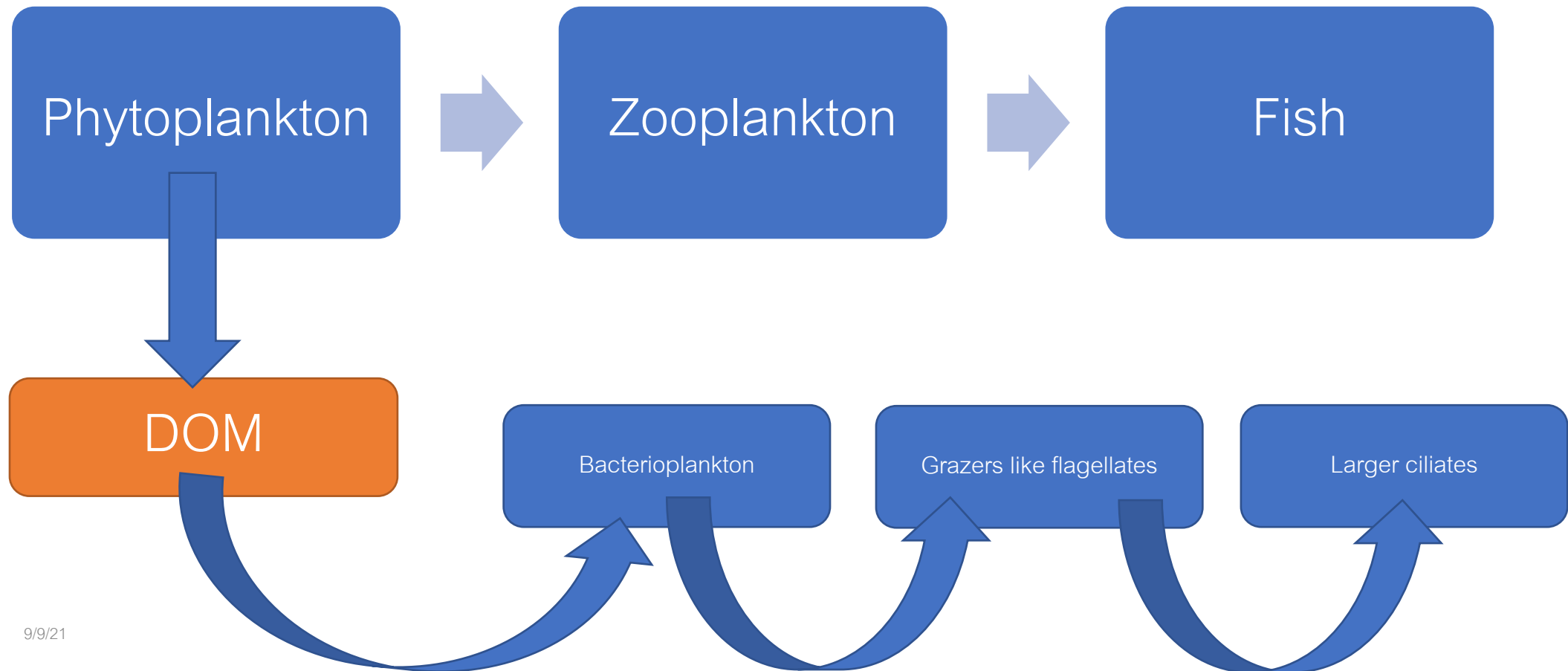


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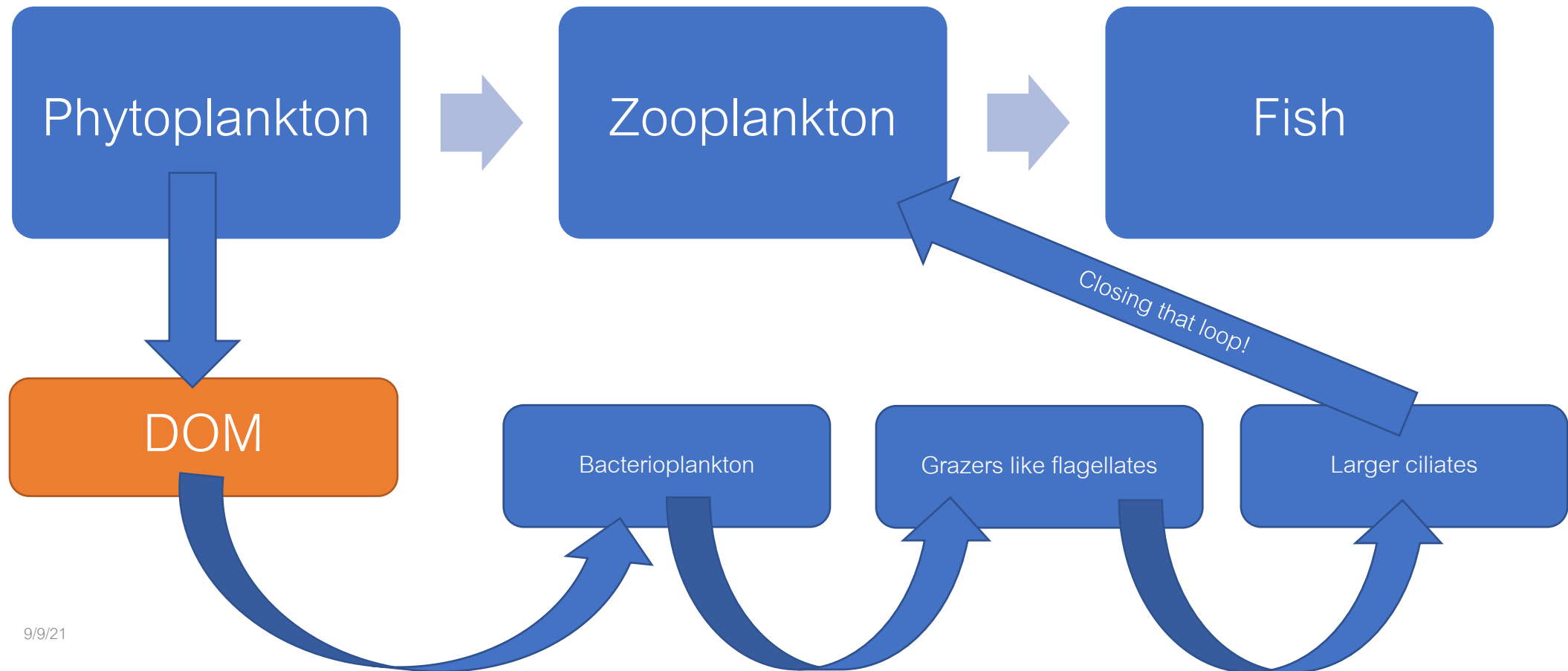




It turns out that this oversimplifies things...



It turns out that this oversimplifies things...



Copepods  
hunting  
phytoplankton

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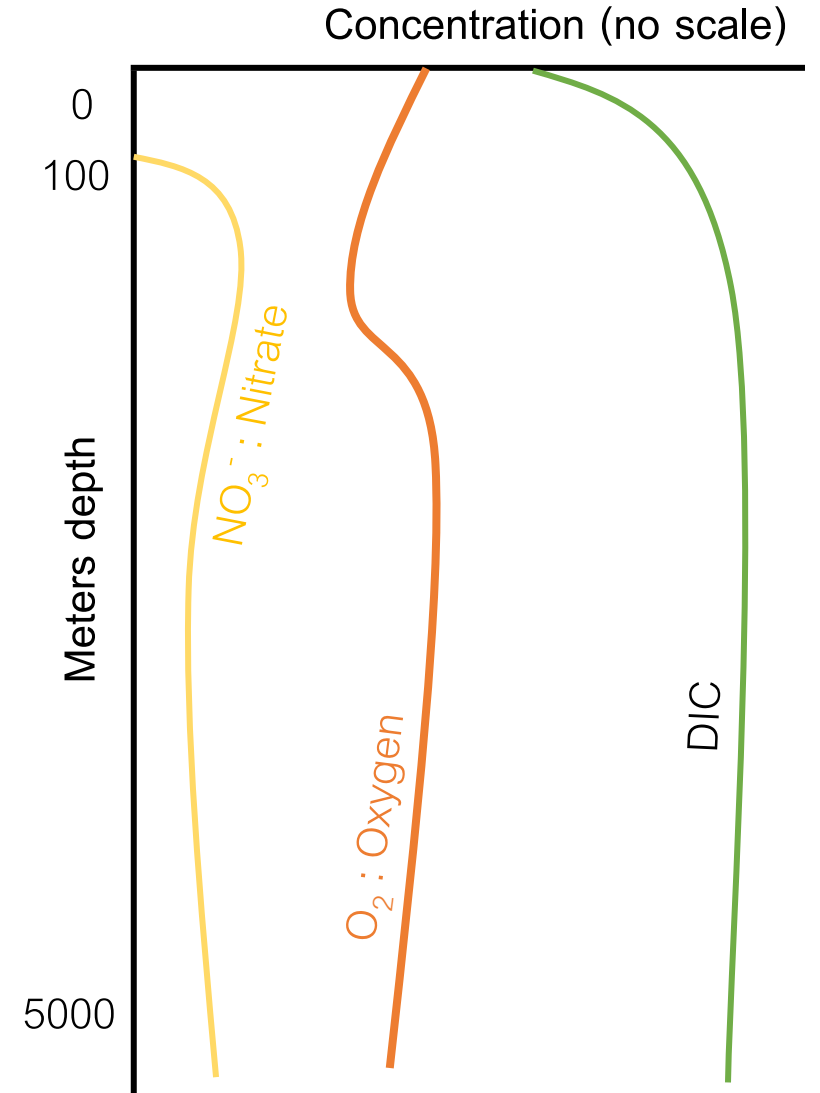
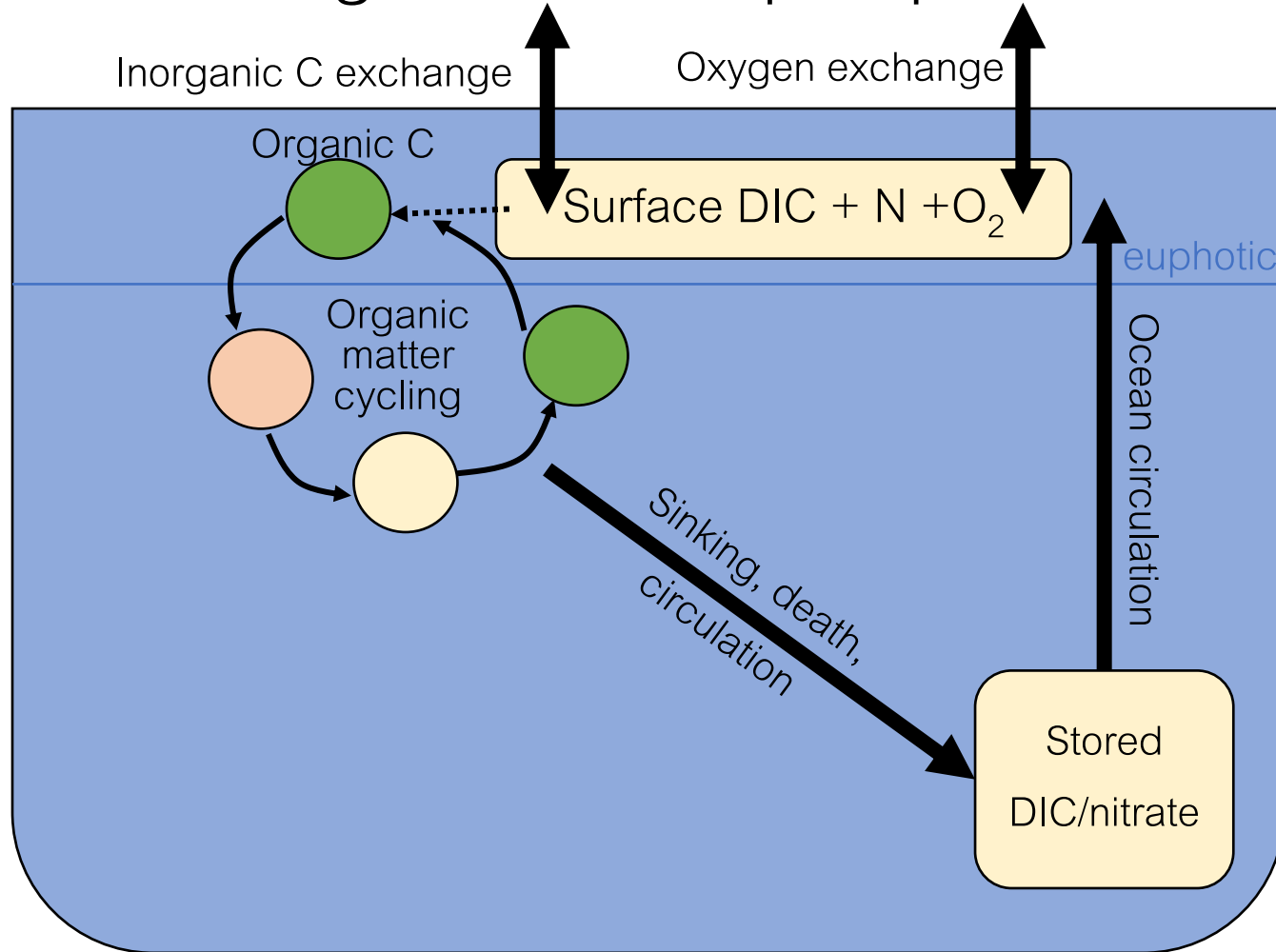


Copepod  
consumes a  
diatom

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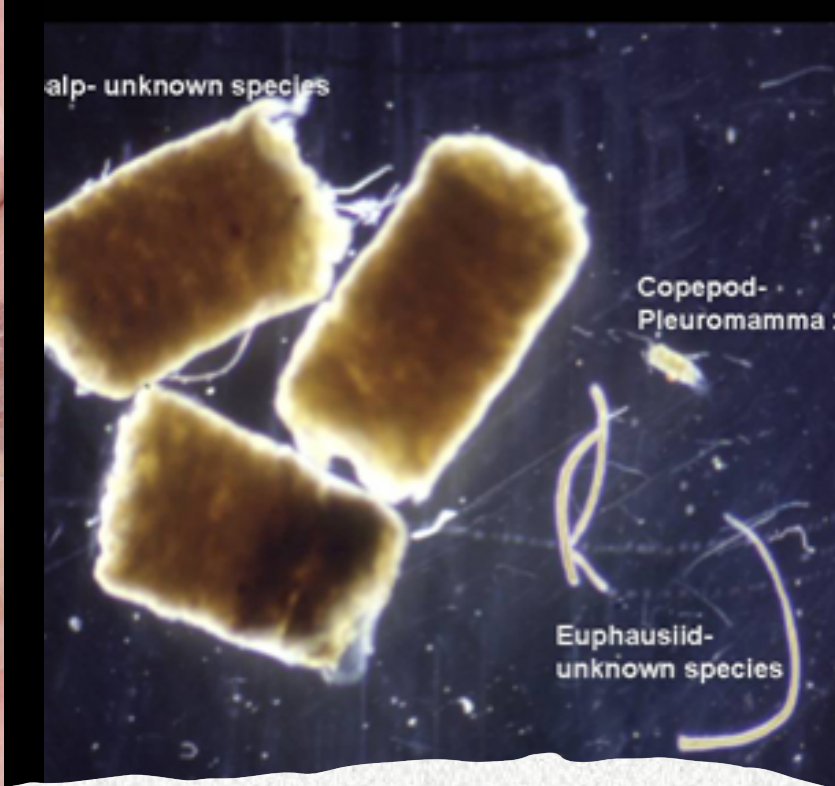


# The biological carbon pump

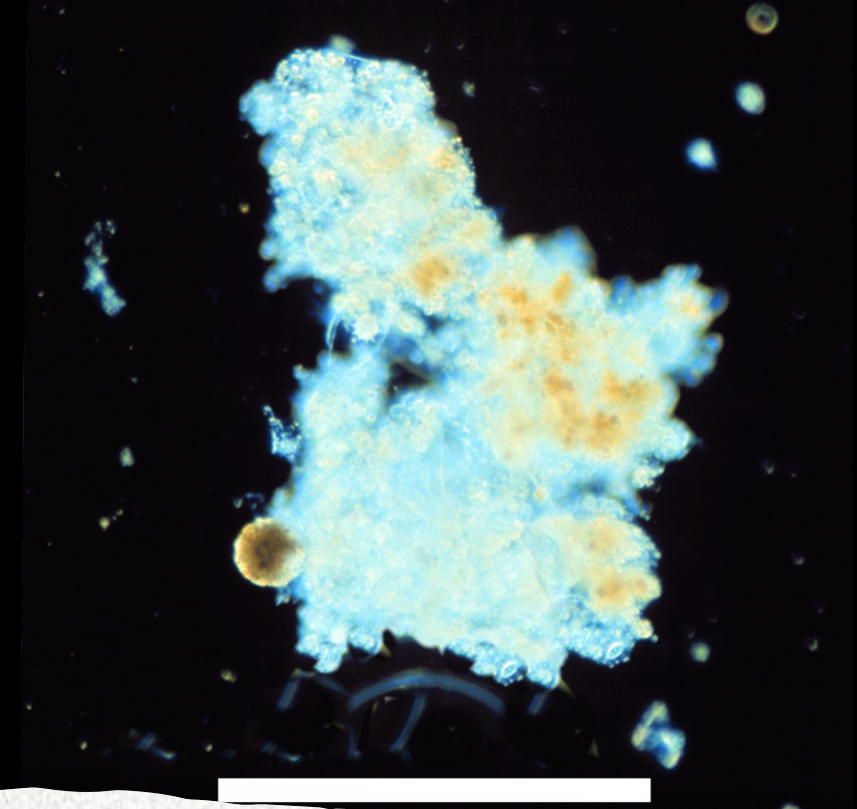




Salp – L. Koren



Zooplankton Fecal Pellets – Debbie Steinberg

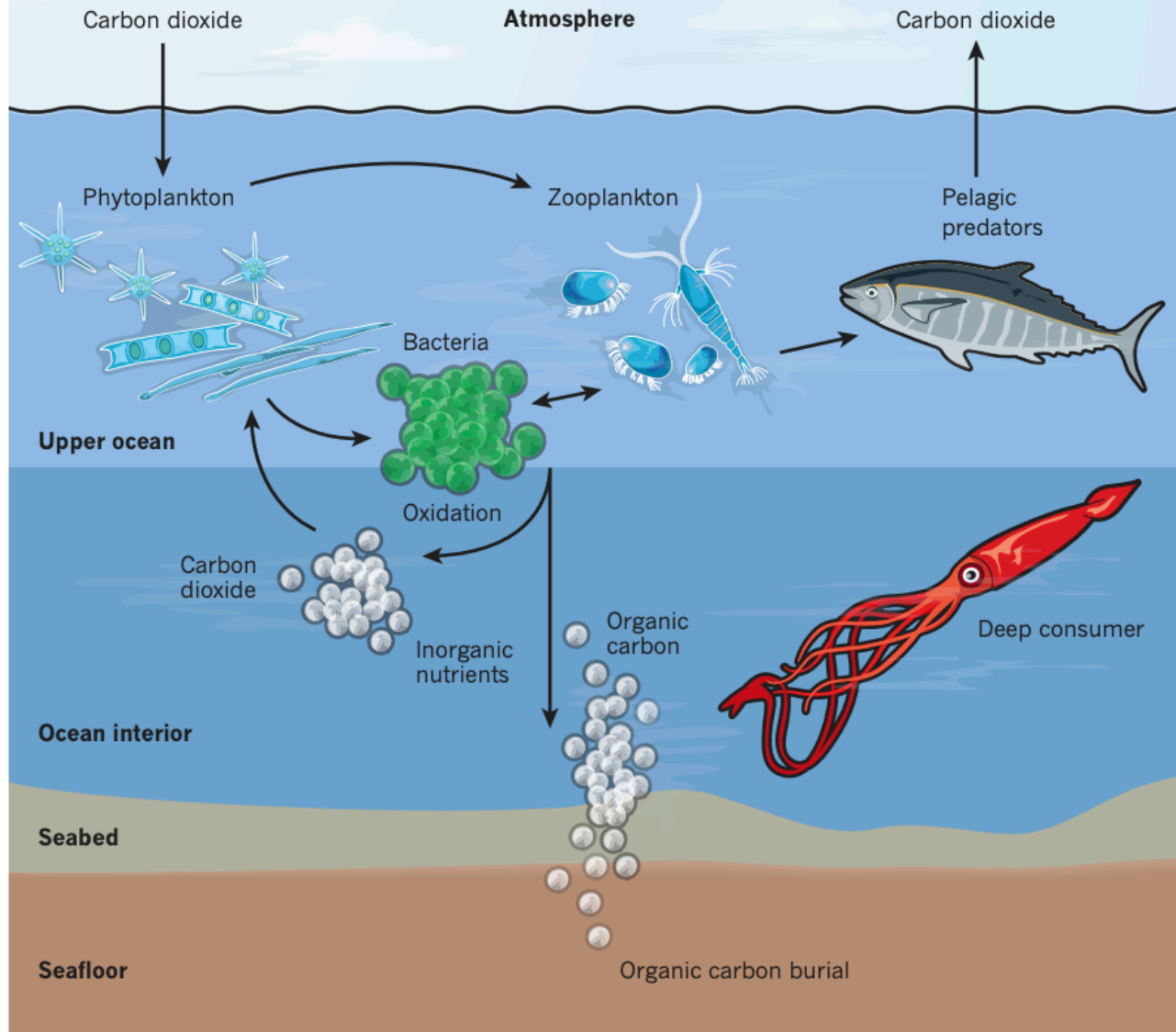


Marine Snow – Richard Lampitt

When particles sink, they export (and potentially bury) these nutrients

## THE BIOLOGICAL PUMP

Phytoplankton drive a biological pump that uses the Sun's energy to move carbon from the atmosphere to the ocean interior, bringing down the atmospheric levels of carbon dioxide.



The biological pump is relevant to global biogeochemical cycles

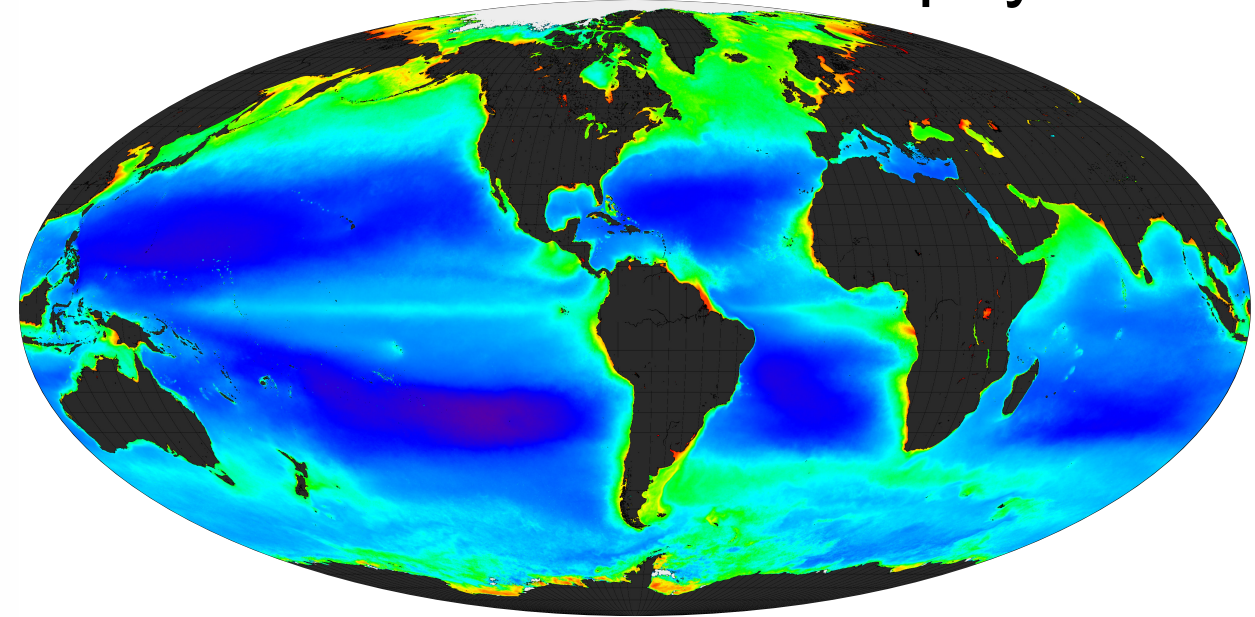
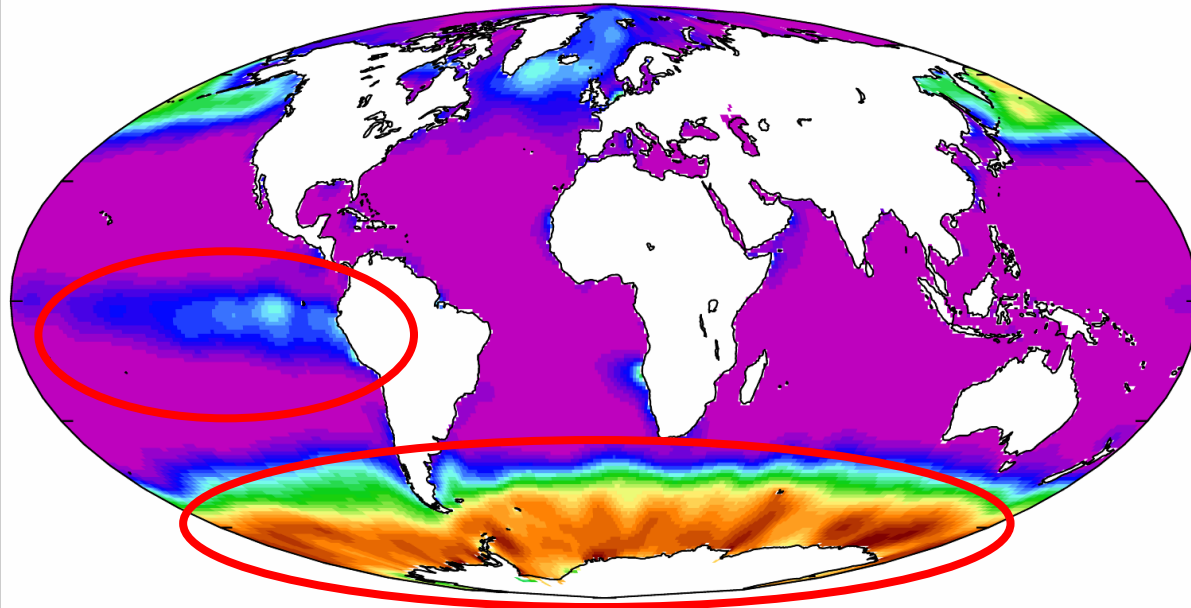
# An exploration of nutrient limitation in the ocean



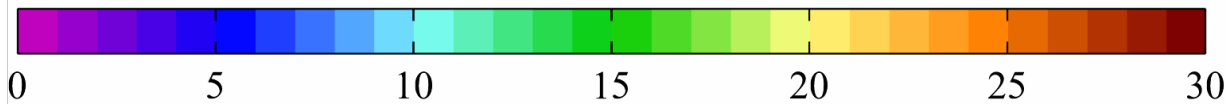
In some areas of the ocean, production stops, but nitrate is still available – “High Nutrient, Low Chlorophyll” regions

Surface nitrate concentration

NASA MODIS chlorophyll

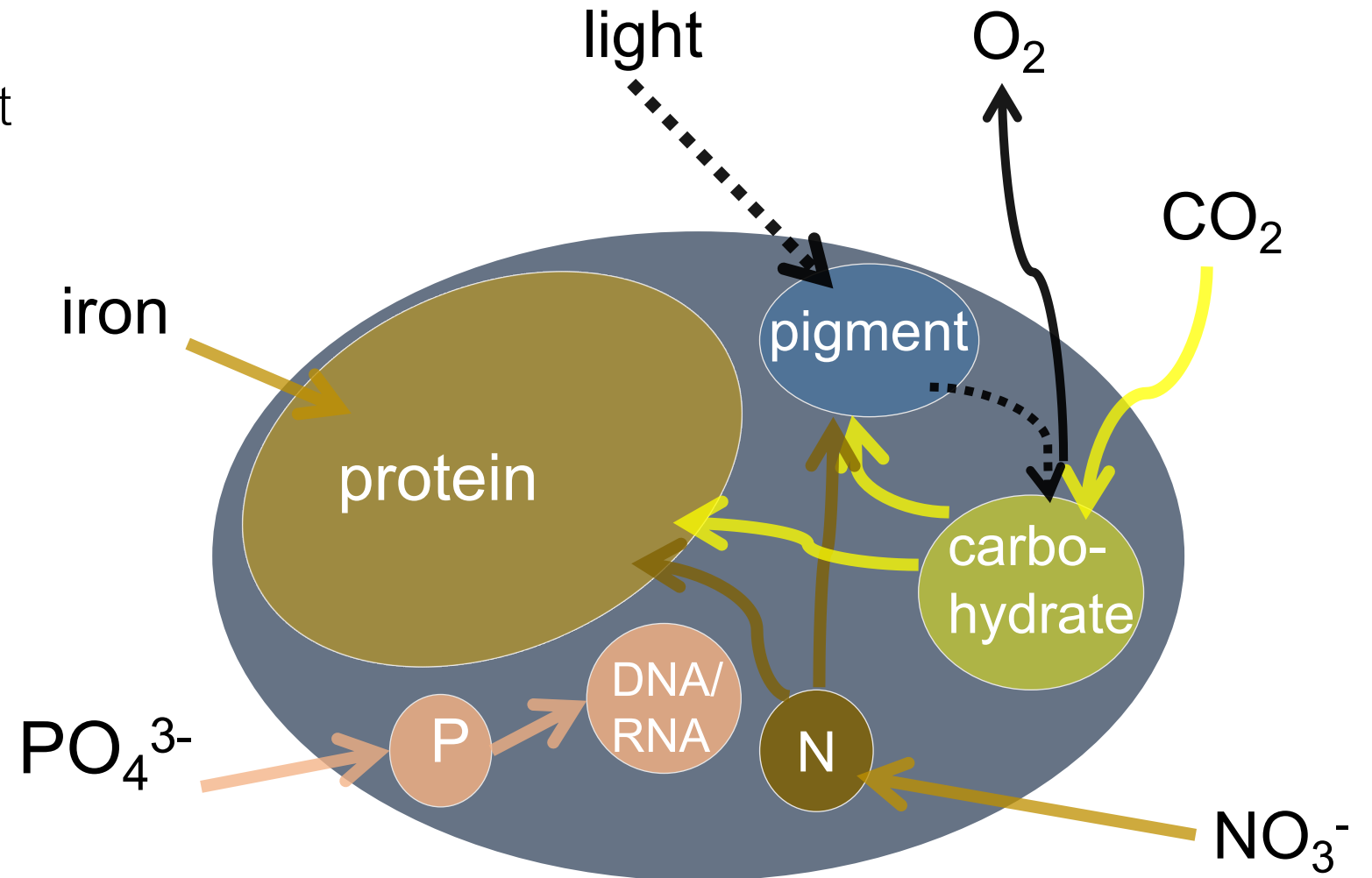


Sea-surface nitrate [ $\text{mmol N m}^{-3}$ ]



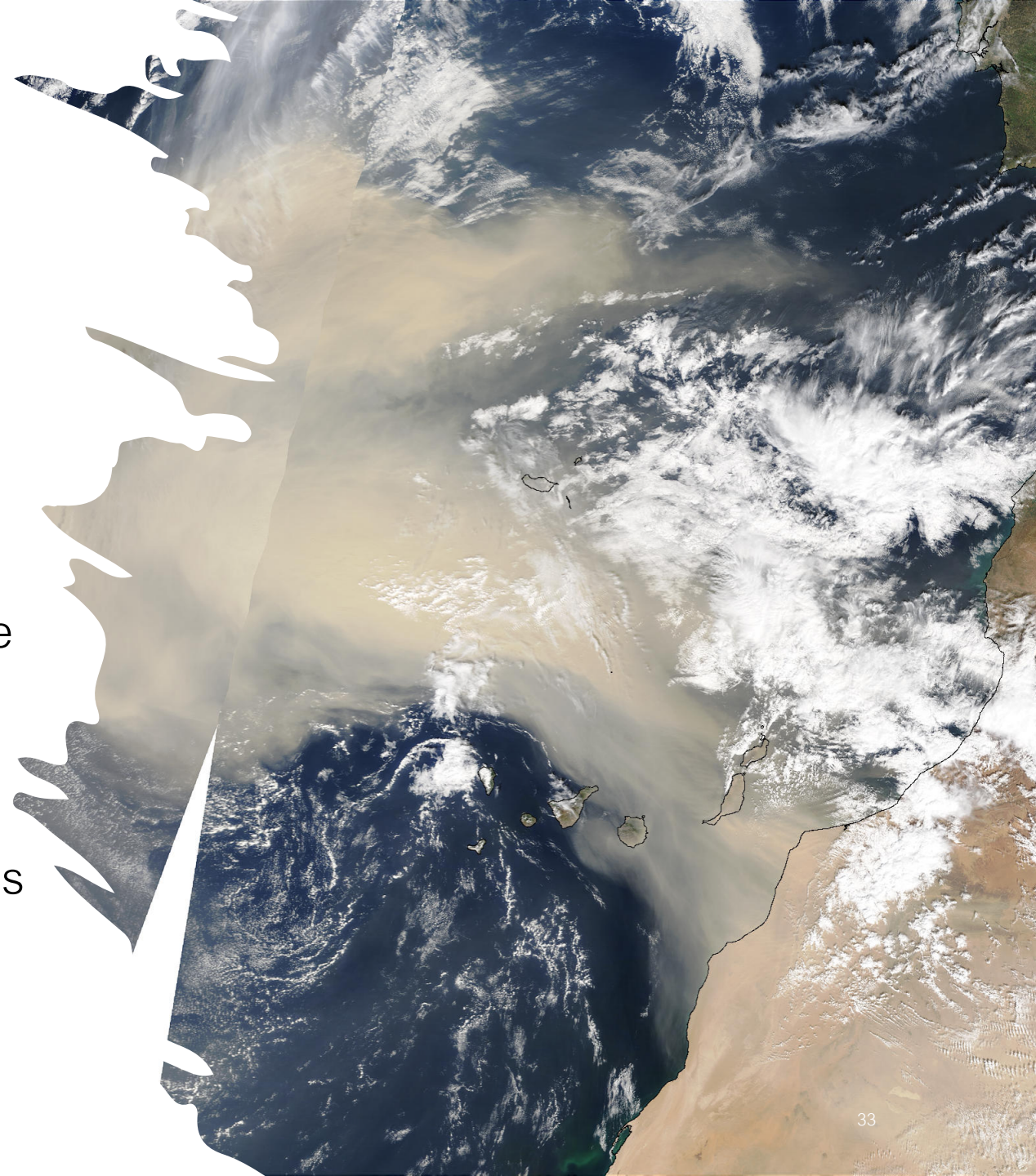
HNLC regions show how the N:P ratio we expect might be off in the oceans

- Some other nutrient must be off from the average Redfield ratio

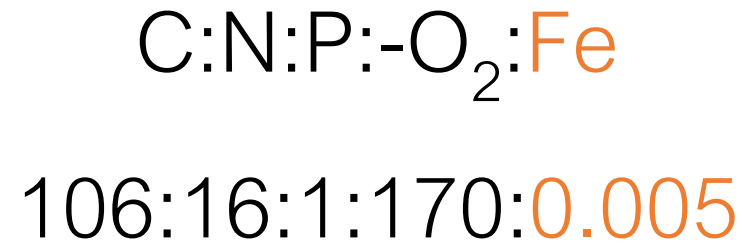


# Trace metals like iron throw a wrench into the plan

- Because iron is a necessary nutrient, but also quite scarce, it is easily scavenged from surface waters
- This means in some upwelling areas, there might not be enough Fe to go around, making nitrate over-abundant
- Dust storms are one of the biggest sources of iron to the ocean, which keeps the Atlantic reasonably well-supplied



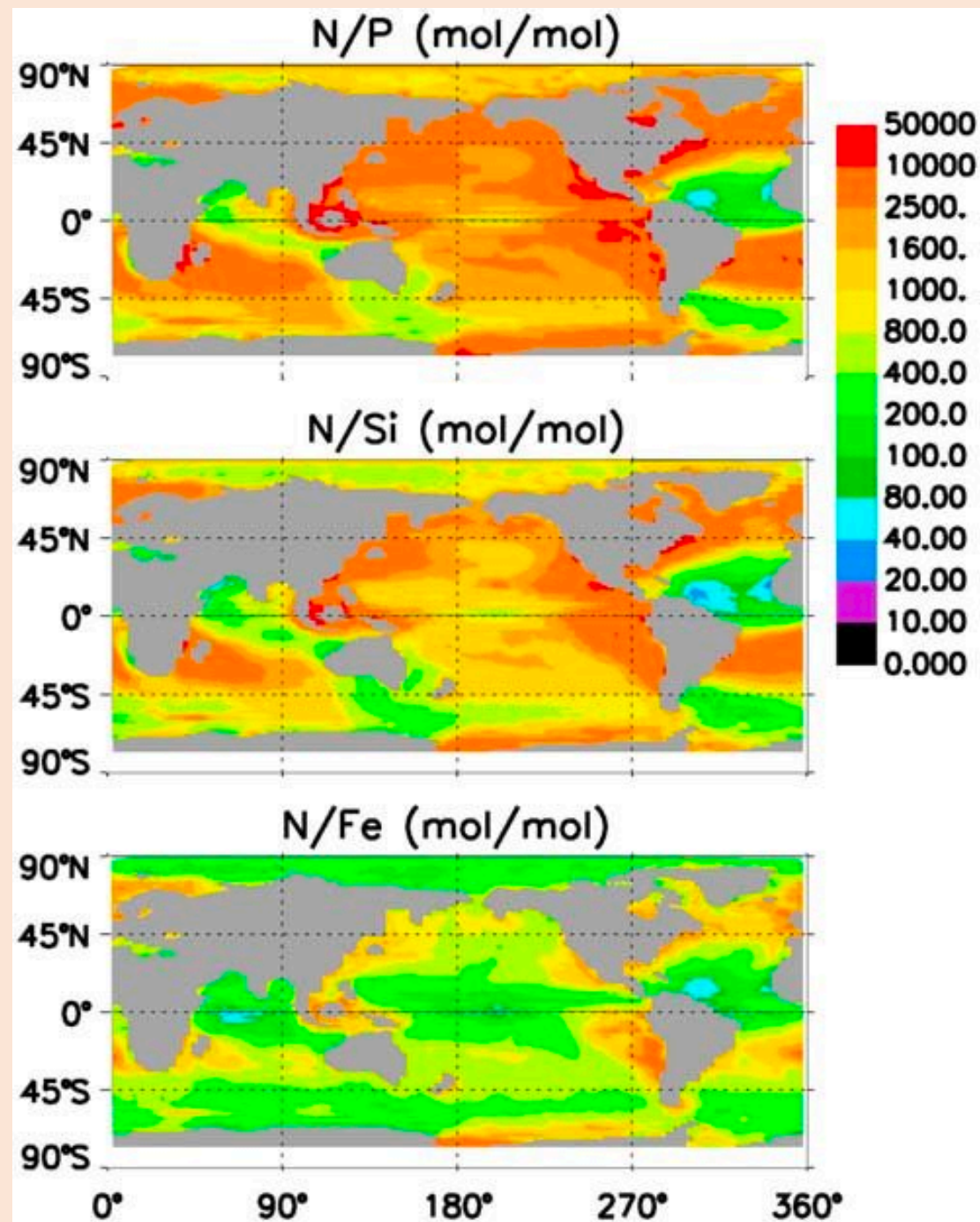
The iron requirement necessitates a new look at the Redfield ratio



For every 16 moles N (one per  $\text{NO}_3^-$ ), 0.005 moles of iron are required. This is a ratio of 3200 N per 1 Fe.

In most of the ocean, N:Fe is much lower than 3200, indicating that we have an excess of Fe relative to N.

In HNLC regions, the pattern is flipped, and the iron availability is too low relative to N.



How can we contextualize  
phytoplankton sizes and functions?

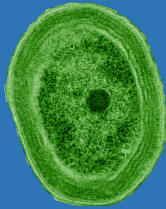
# The mind-boggling scales of (marine) phytoplankton



# The mind-boggling scales of (marine) phytoplankton



**Prochlorococcus**



0.7 microns



(20 inches)

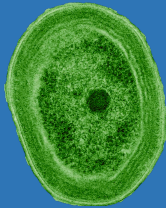


# The mind-boggling scales of (marine) phytoplankton

5-20 microns  
(10x bigger than 0.5)

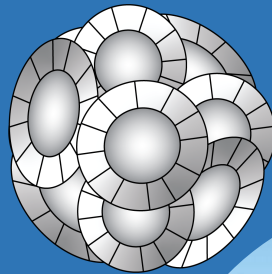


**Prochlorococcus**



0.7 microns

***Emiliana huxleyi***



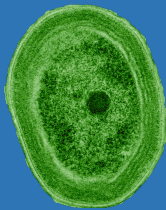
(20 inches)

# The mind-boggling scales of (marine) phytoplankton

*Karenia brevis*



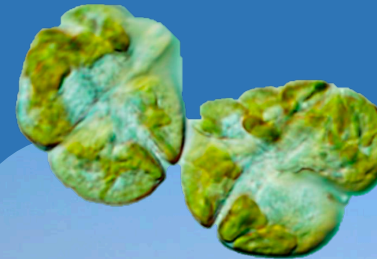
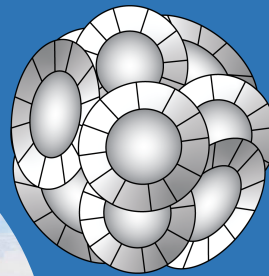
**Prochlorococcus**



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*Emiliana huxleyi*



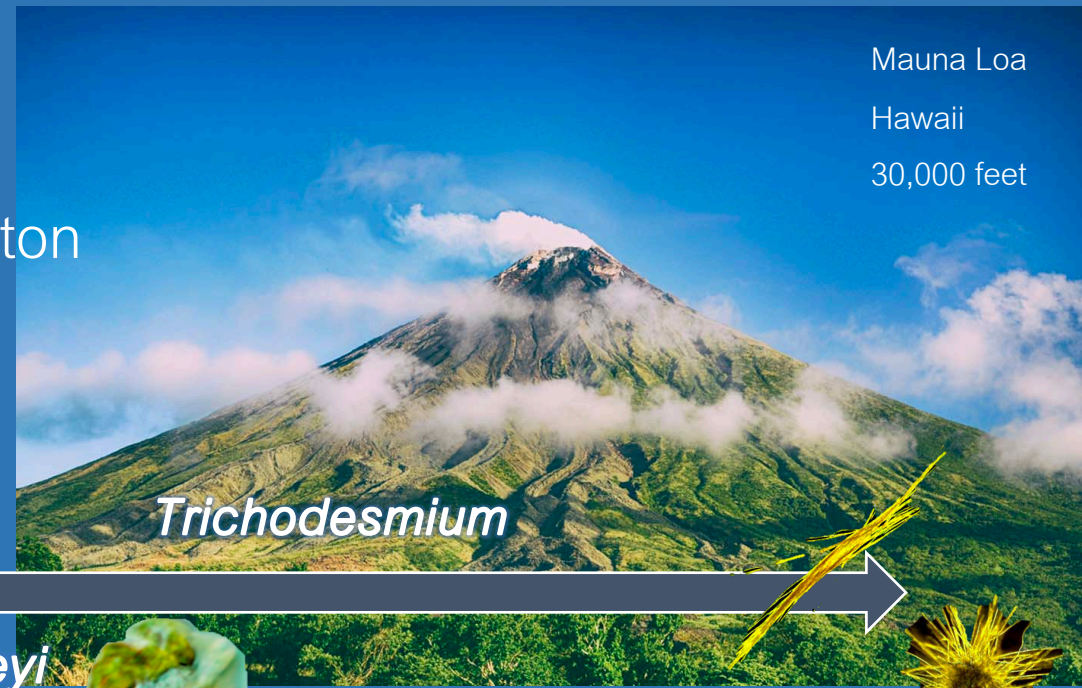
(20 inches)



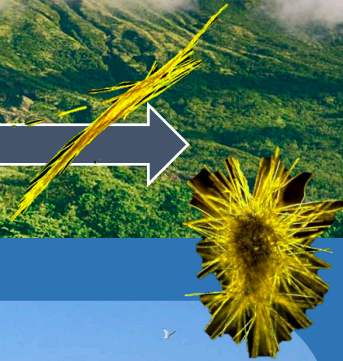
Ha'Penny Bridge  
Dublin, Ireland  
(135 feet)

# The mind-boggling scales of (marine) phytoplankton

Mauna Loa  
Hawaii  
30,000 feet



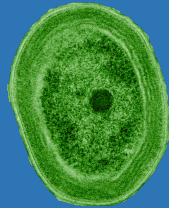
*Trichodesmium*



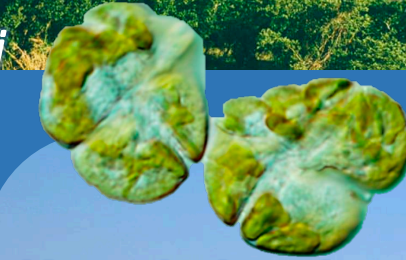
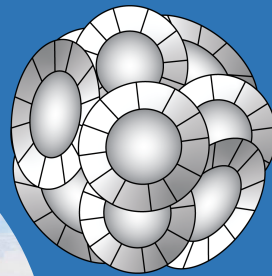
*Prochlorococcus*

5-20 microns  
(10x bigger than 0.5)

*Emiliana huxleyi*



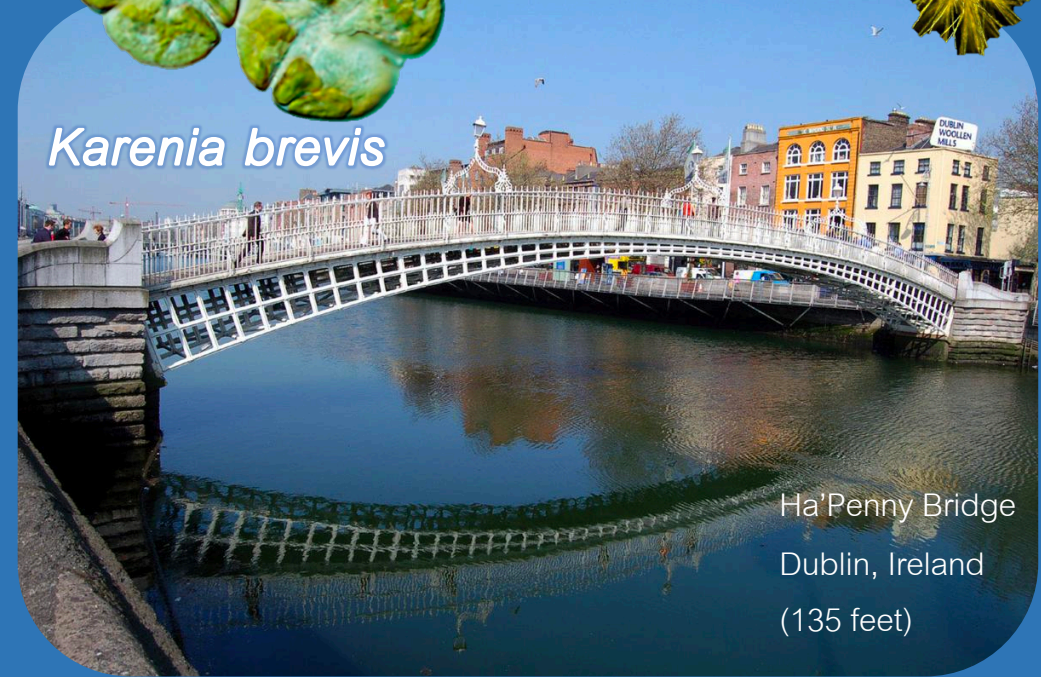
0.7 microns



*Karenia brevis*

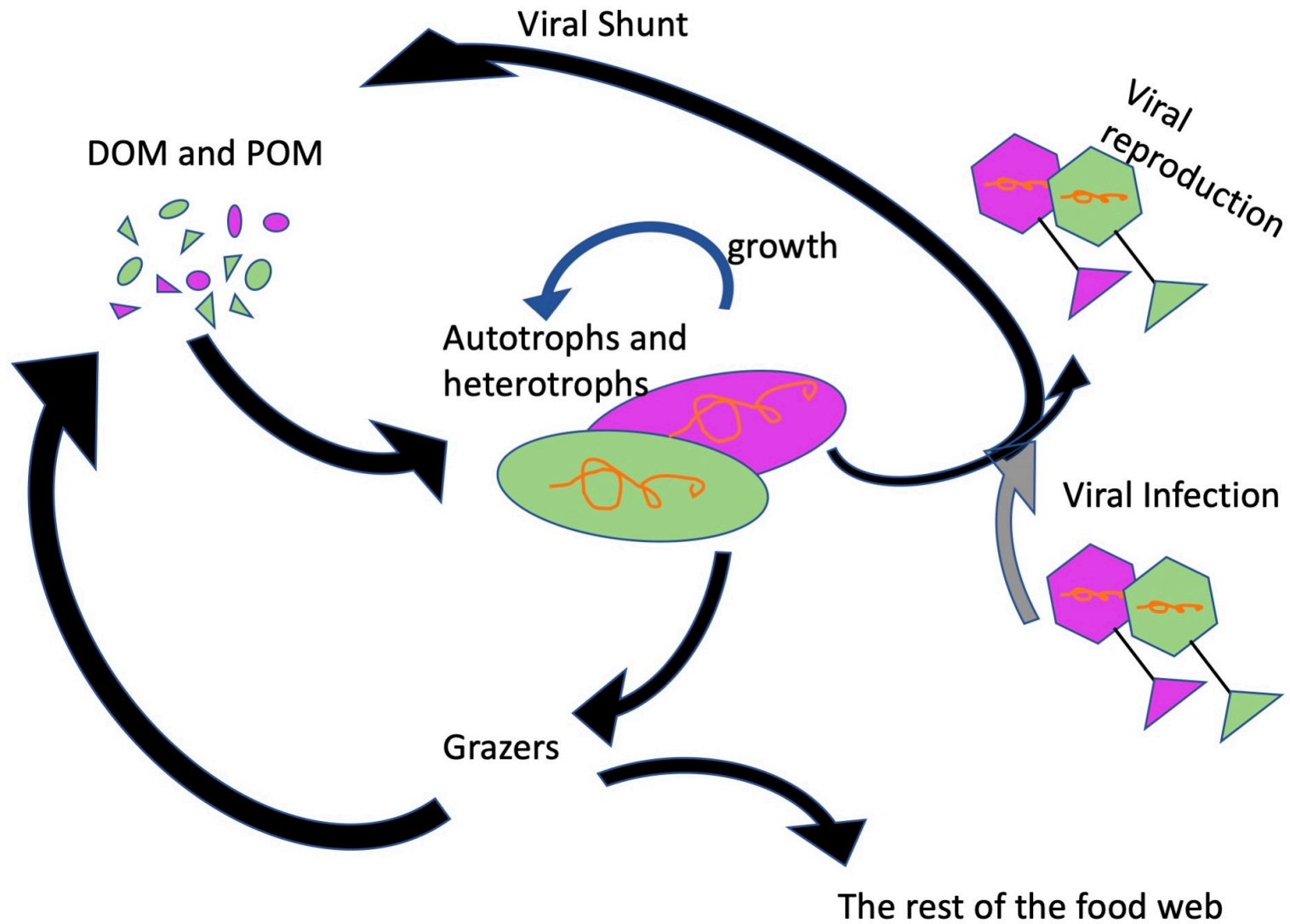


(20 inches)

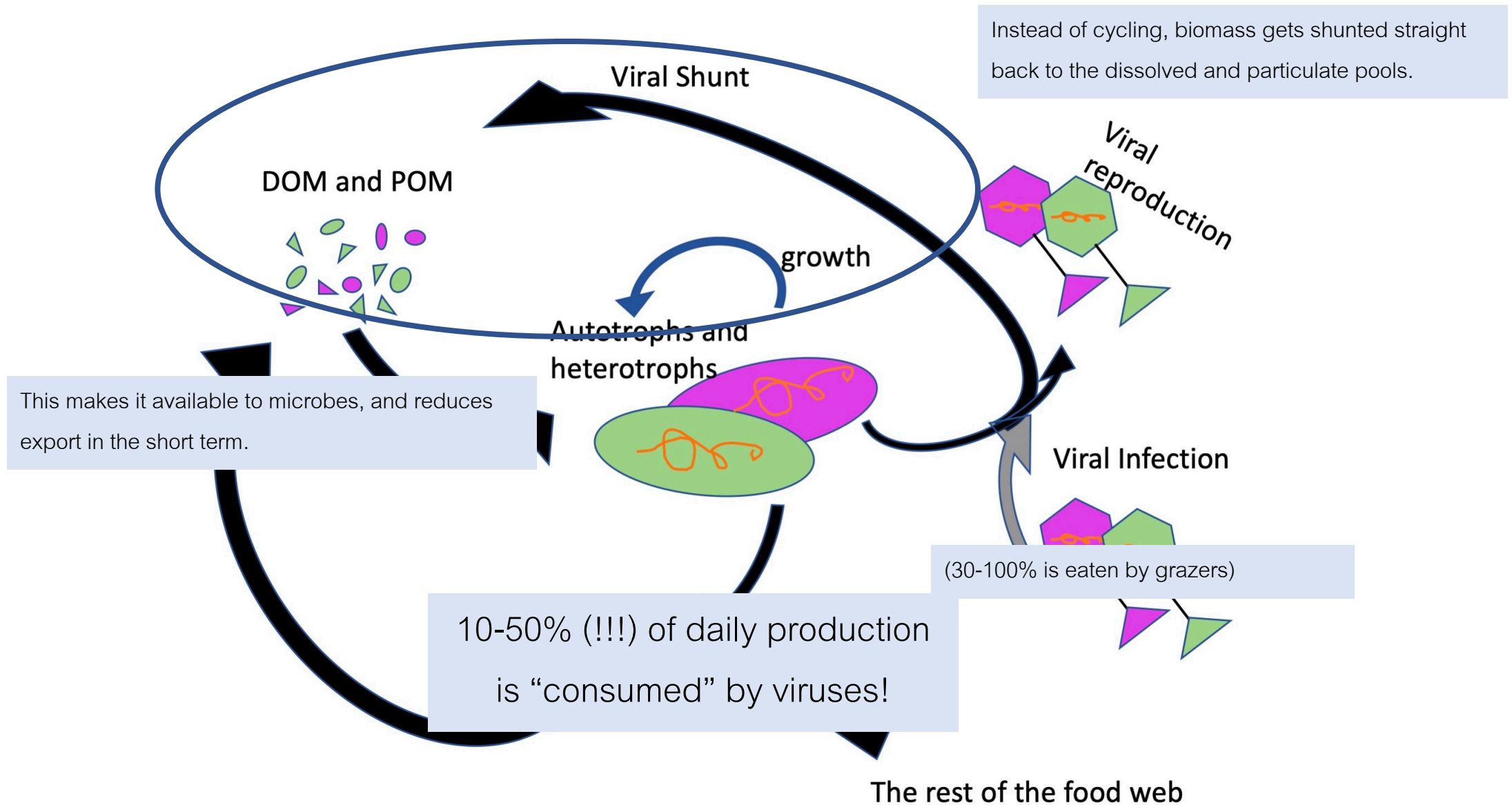


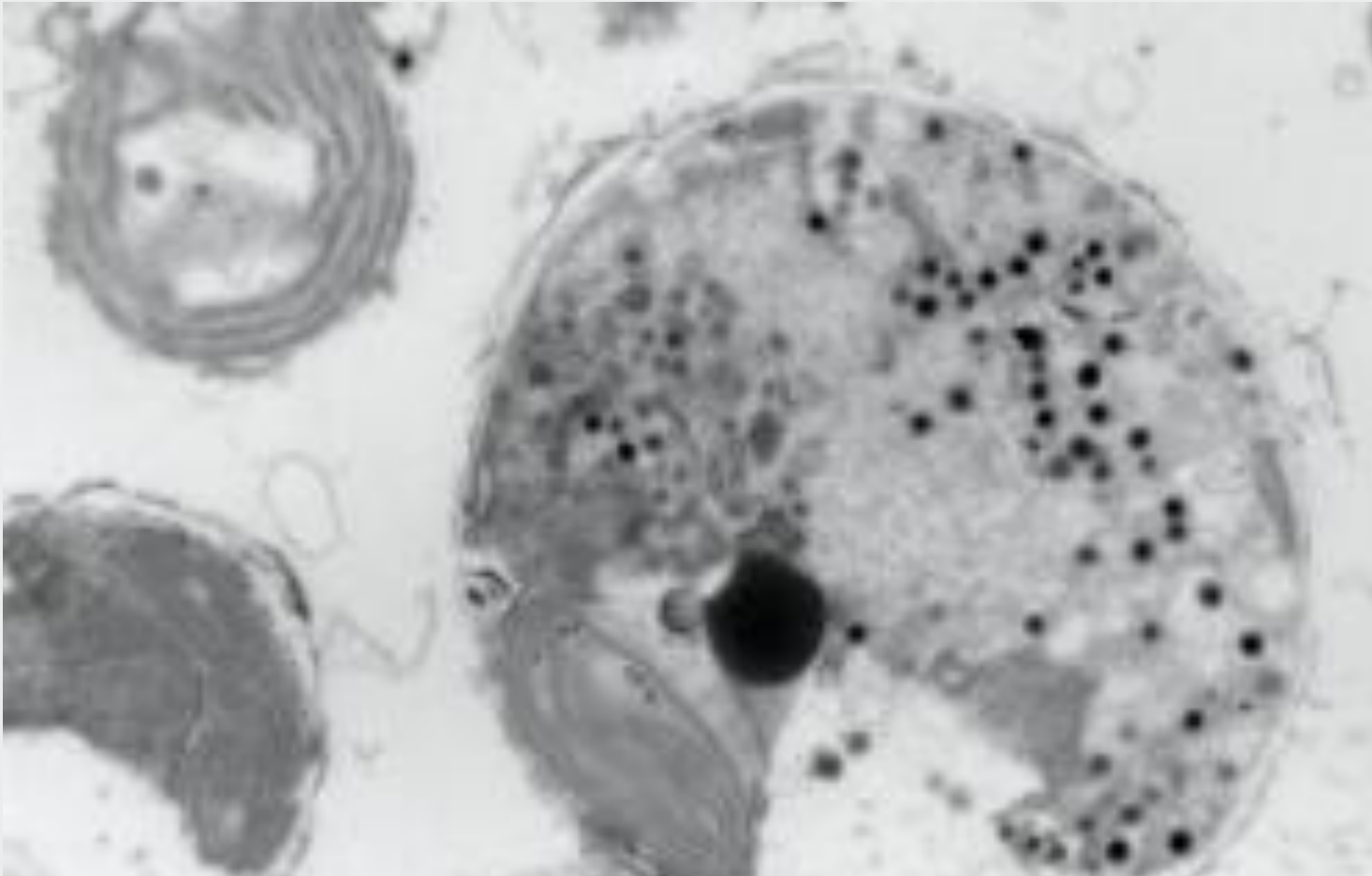
Ha'Penny Bridge  
Dublin, Ireland  
(135 feet)

# The microbial loop is further complicated by viral activity



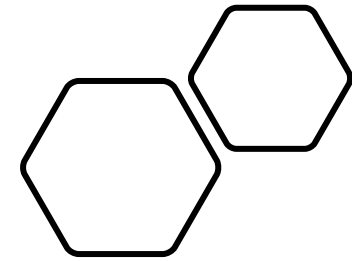
# The microbial loop is further complicated by viral activity






*E. huxleyi* infected by virus (black spots)

1  $\mu\text{m}$



A dense, overlapping collection of various phytoplankton species, including diatoms, cyanobacteria, and other microorganisms, shown in various colors and shapes. The background is a light, textured surface.

Phytoplankton are too diverse to adequately lump into categories

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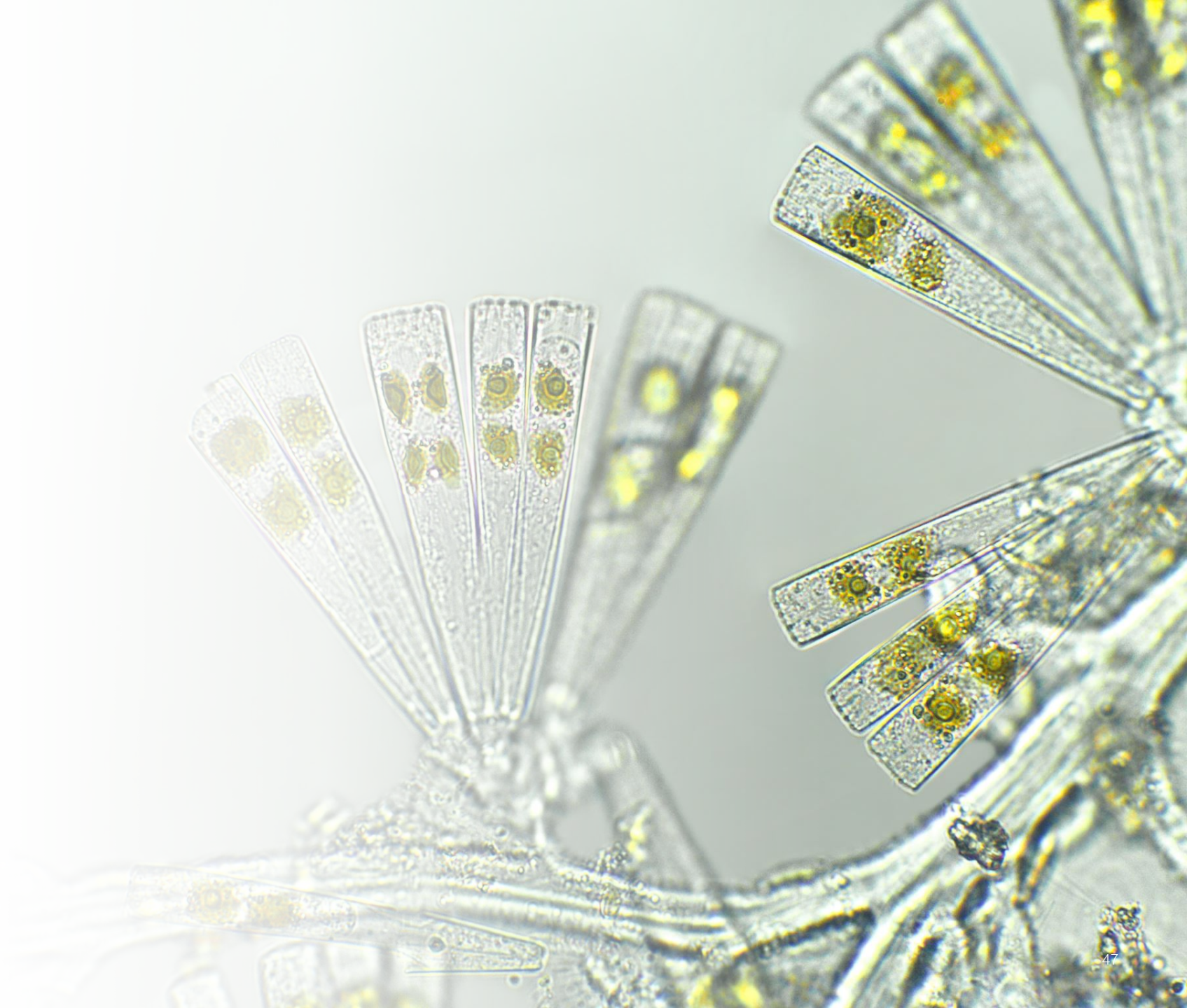
- Thousands of phytoplankton can be uniquely morphologically identified
- These can be classified into taxonomic groups which typically correspond to biogeochemical function

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## Major functional classes of phytoplankton

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- Calcifiers
- Nitrogen fixers
- Silicifiers
- DMS producers

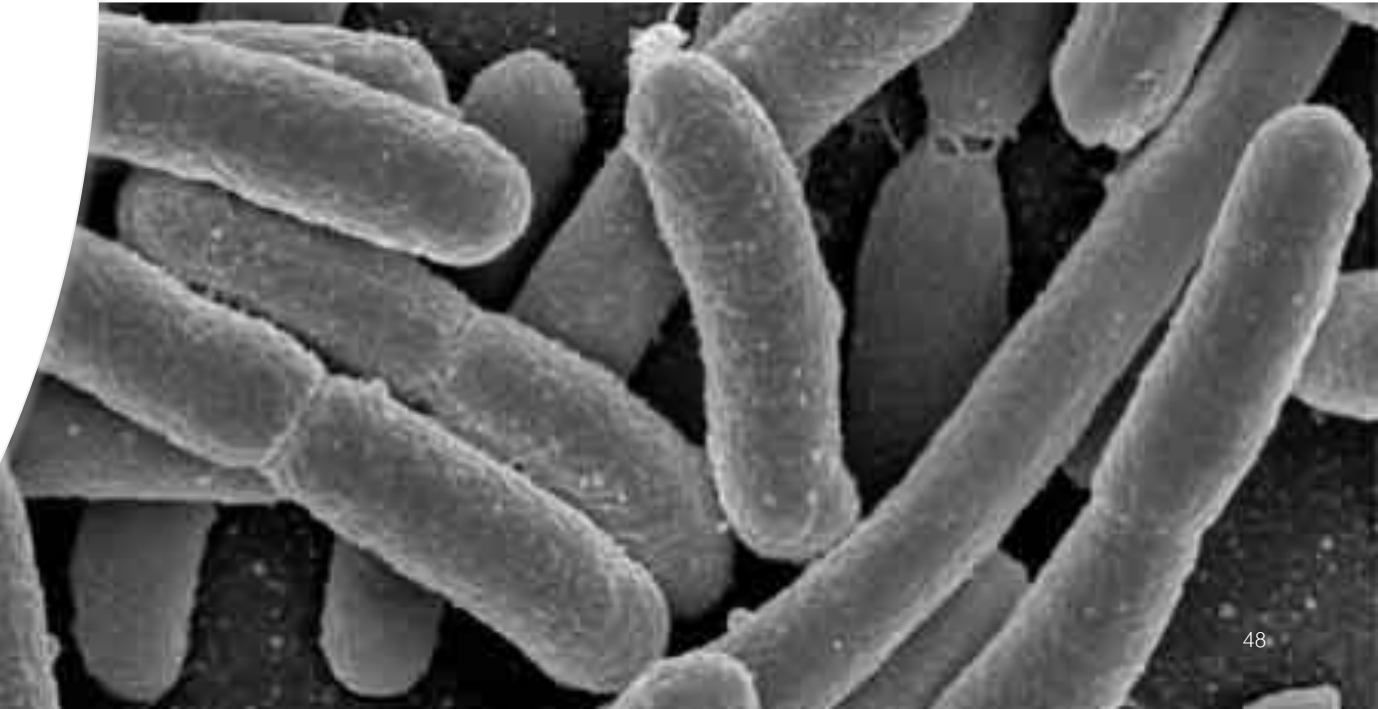
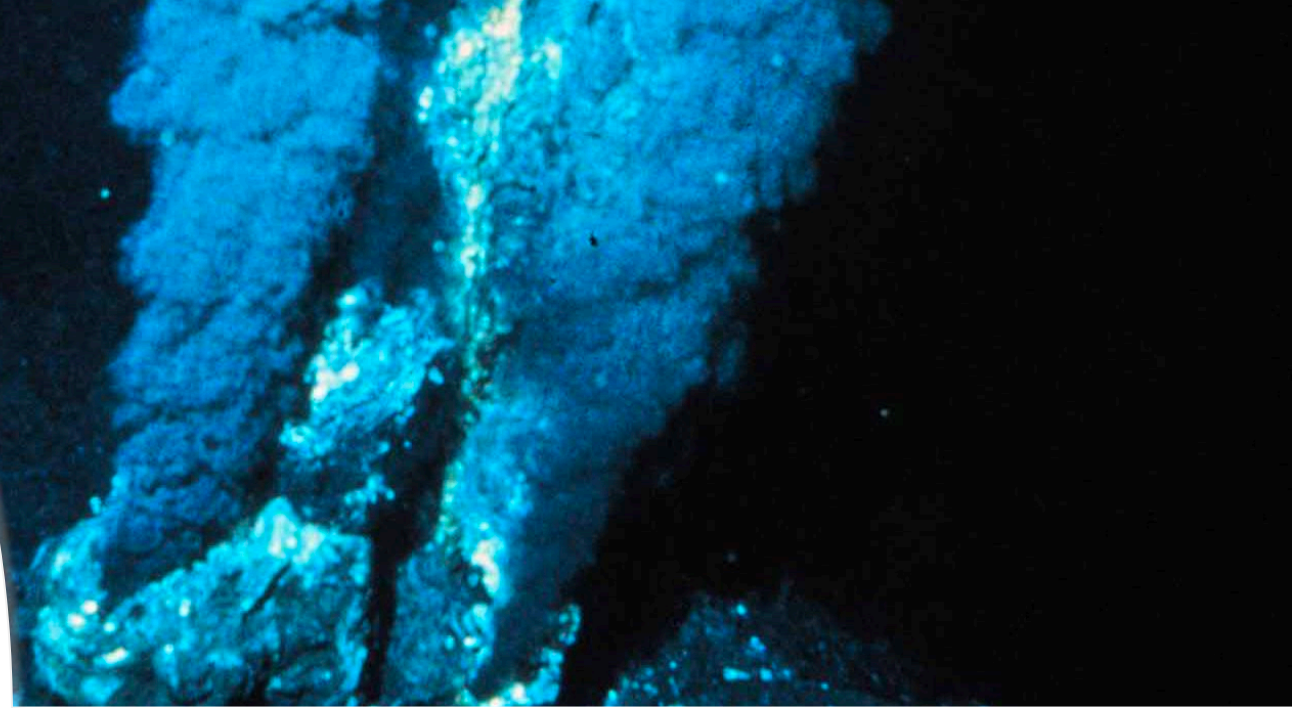




## Other ecologically-relevant microbes

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- Traditional denitrifying + nitrifying bacteria + archaea
- Anammox-involved bacteria + archaea
  - Also in hydrothermal vents – whole other topic!!



# Pico-cyanobacteria

# Pico-cyanobacteria



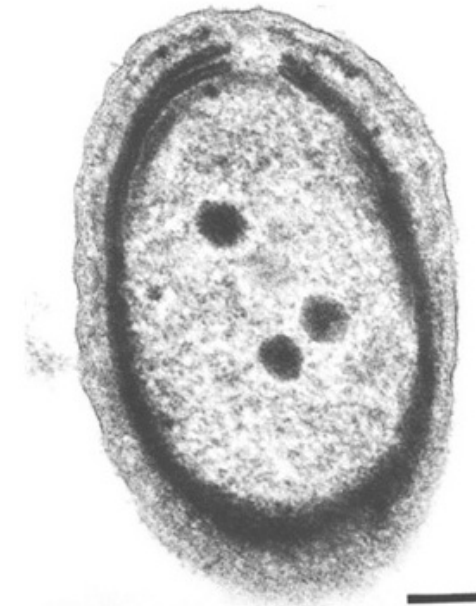
Prokaryotic (not true algae); genome ~1.7-9 Mbp



Smallest photo-autotrophs in existence, in particular  
Prochlorococcus (<1  $\mu\text{m}$  Effective Spherical Diameter/ESD)



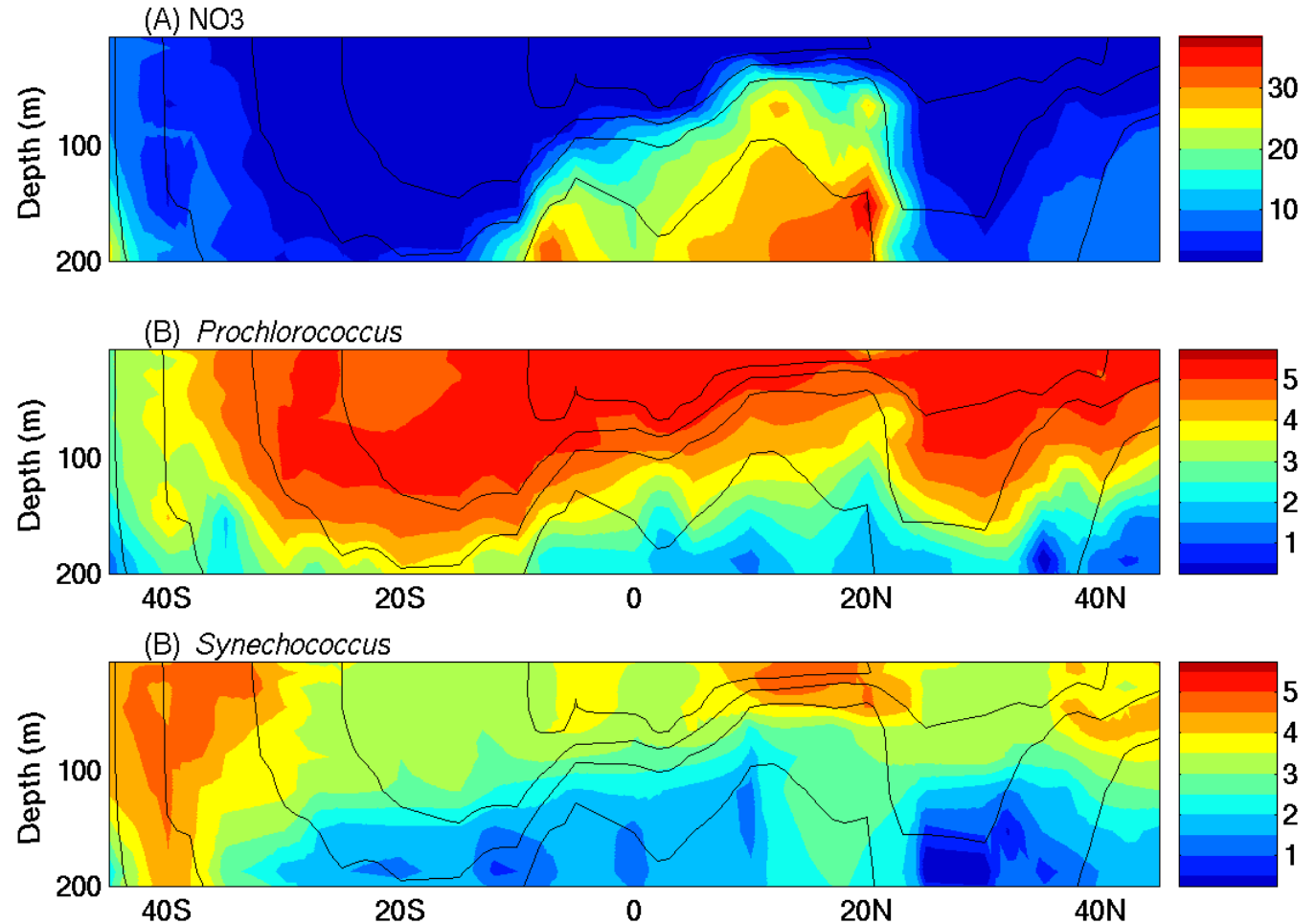
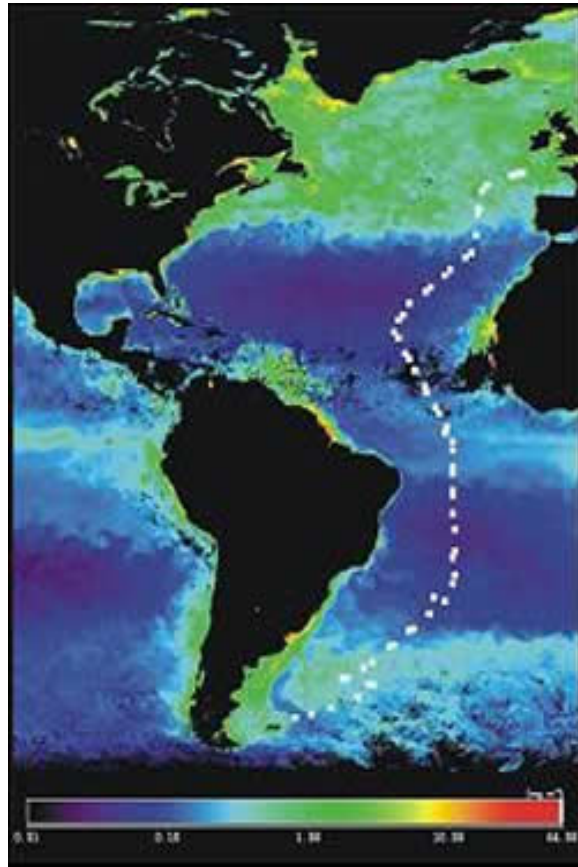
As much as half of the biomass in subtropical water



0.1 micron

*Prochlorococcus*:  
image C. Ting

# Low-nutrient subtropical gyres are picocyanobacterial stomping grounds



Phytoplankton:  $\log(\text{cells ml}^{-1})$   
Nitrate ( $\mu\text{mol kg}^{-1}$ )

Johnson et al, Science (2006)

# Coccolithophores

# Some example calcifiers



*Emiliana huxleyi*

Haptophyte

Associated with EhV, large viruses that

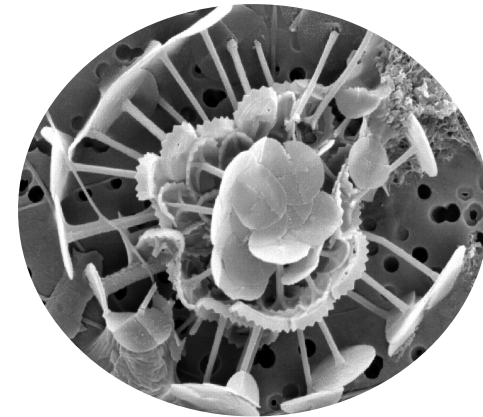
collapse blooms

Coccolithophore poster child



*Calcidiscus*

*leptoporus*



*Papposphaera*

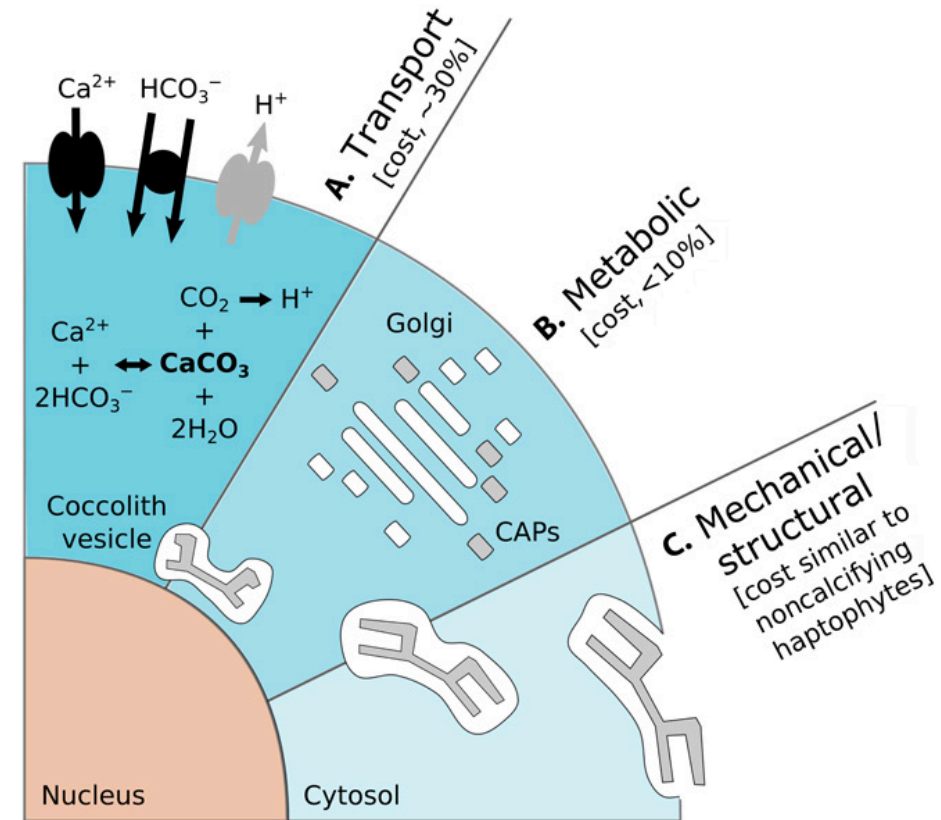
*lepida*

# Coccolith formation in coccolithophores (Allison Taylor, UNC)



# Calcification: an enigmatic process

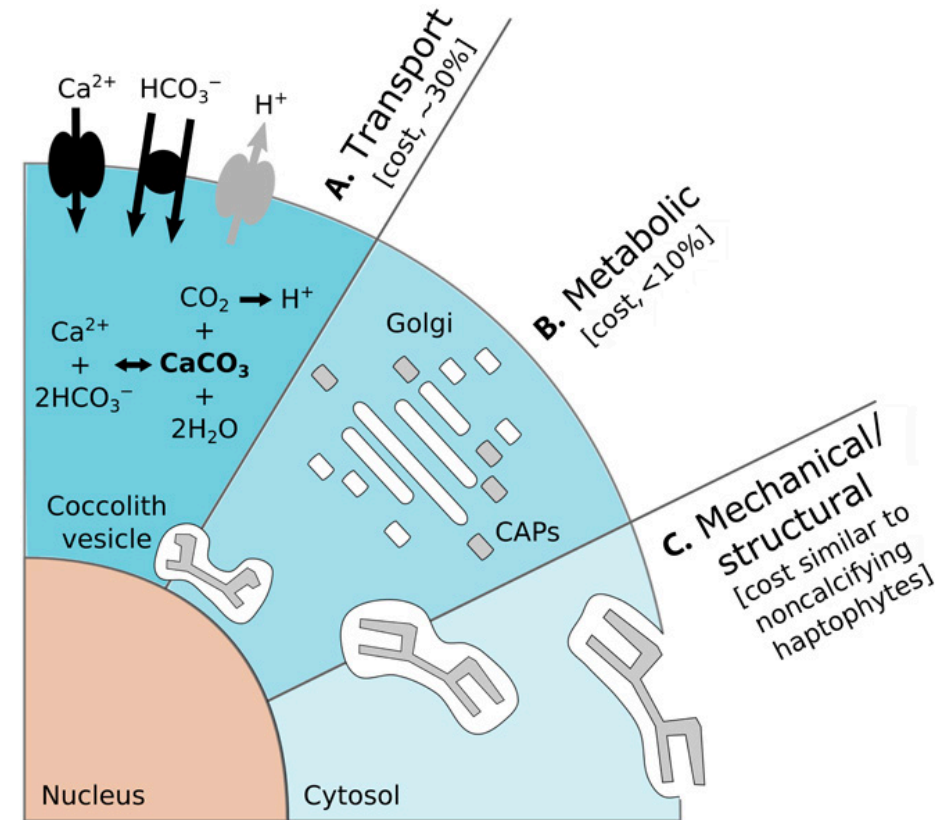
- Calcifiers are abundant, and the calcification process is complex with respect to global  $\text{CO}_2$



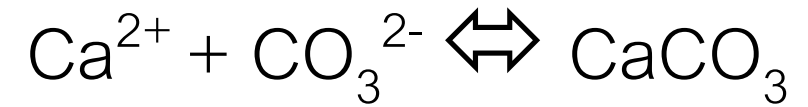


# Calcification: an enigmatic process

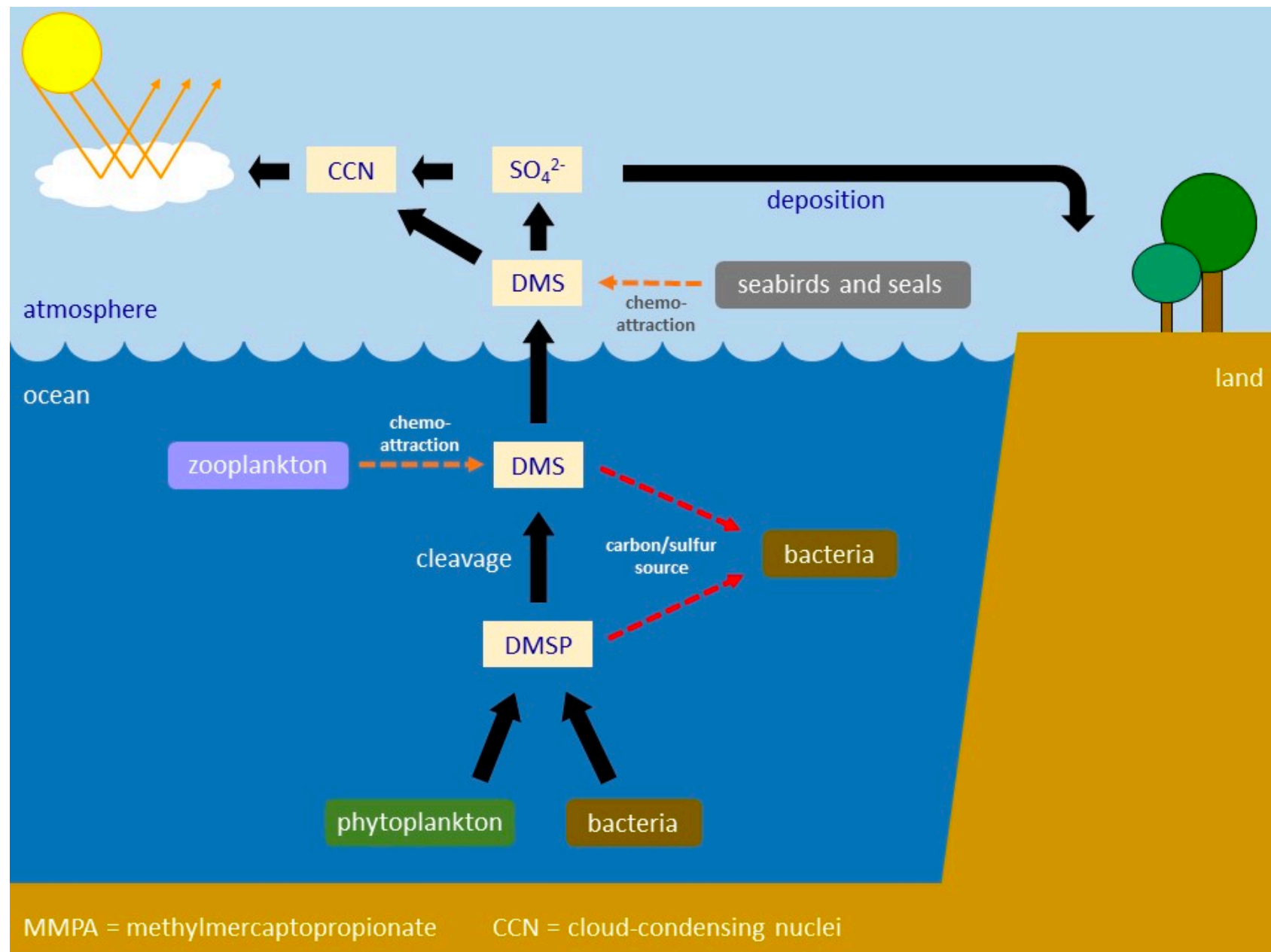
- No one quite knows how calcifiers *afford* the energetic costs of calcification



Calcification can result in large-scale calcite formations after blooms + deposits



DMS  
production in  
*Emiliana  
huxleyi*

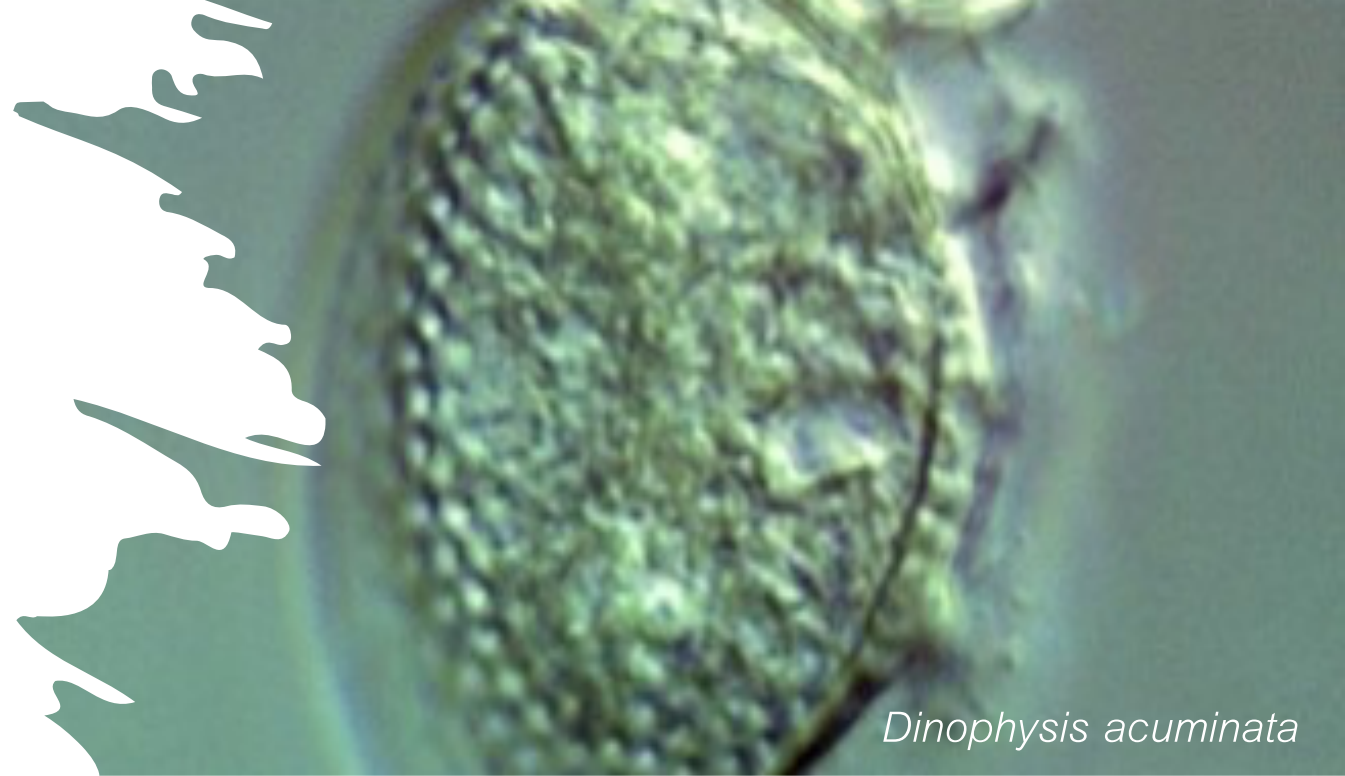


Curson et al. 2018

# Dinoflagellates

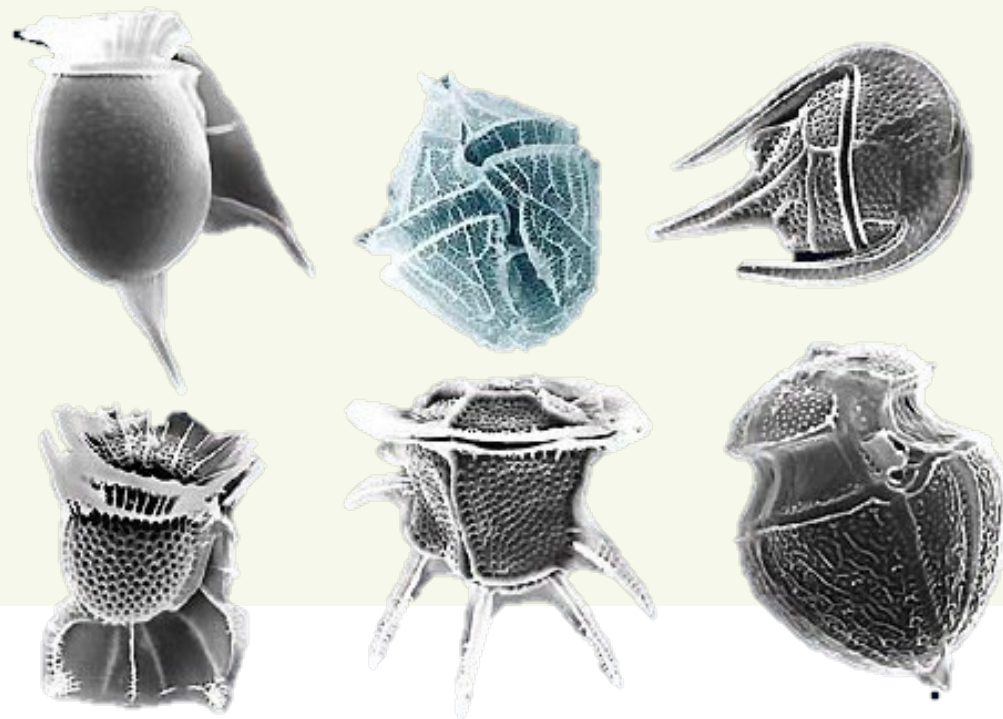
# Dinoflagellate key traits

- 10s to 100s of microns
- Have flagella, but no frustule/mineral
- Grow relatively slowly and often **mixotrophic**



# Toxic bloom-formers

- This is one of the most diverse groups of phytoplankton around
- Includes red tide species *Karenia brevis*
- Also responsible for a lot of surface **bioluminescence**



# Feeding in dinoflagellates

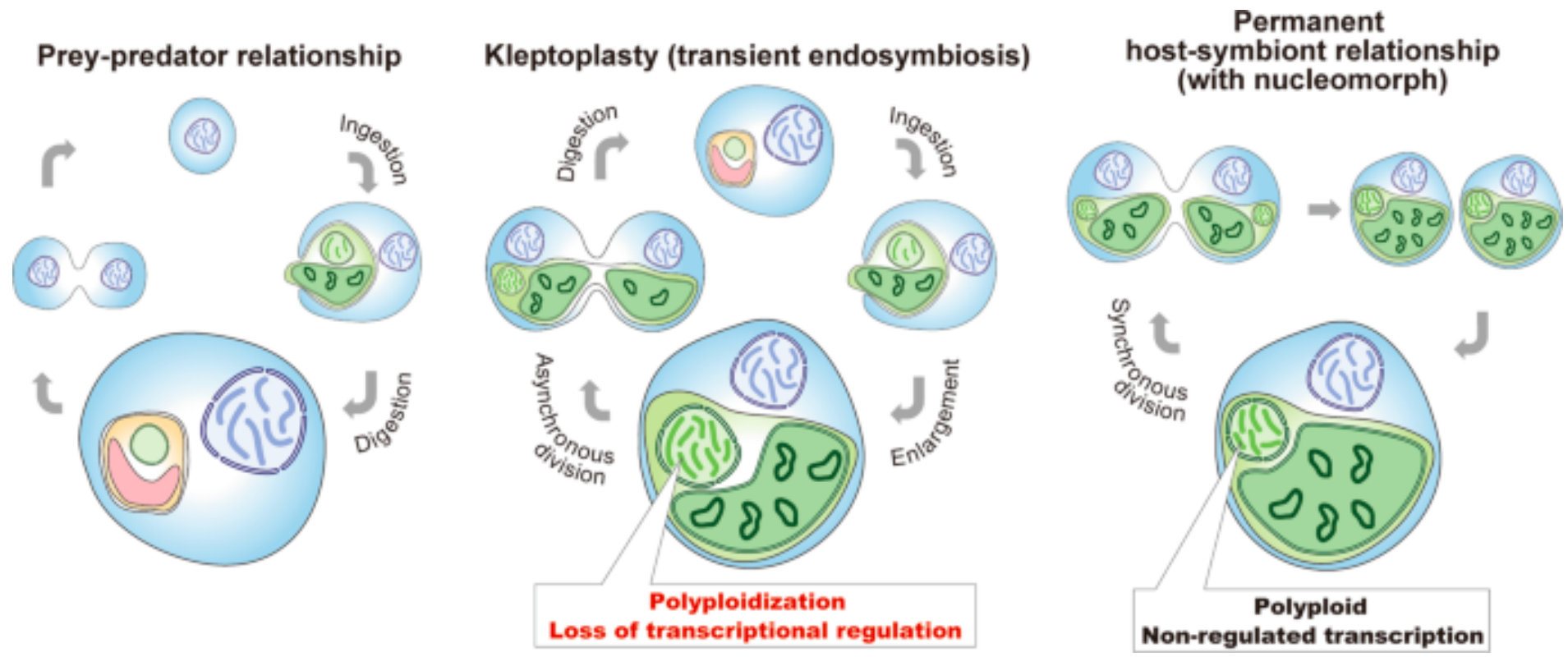


# Dinoflagellates have complex evolutionary histories

- Many phytoplankton gained photosynthetic ability through acquiring **plastids**
- This involves an endosymbiotic event, in which a photosynthetic bacterium is engulfed and eventually it becomes an organelle rather than a separate organism
- In dinoflagellates this has happened *multiple* times independently



# Kleptoplasty – piecewise motion towards photosynthetic ability

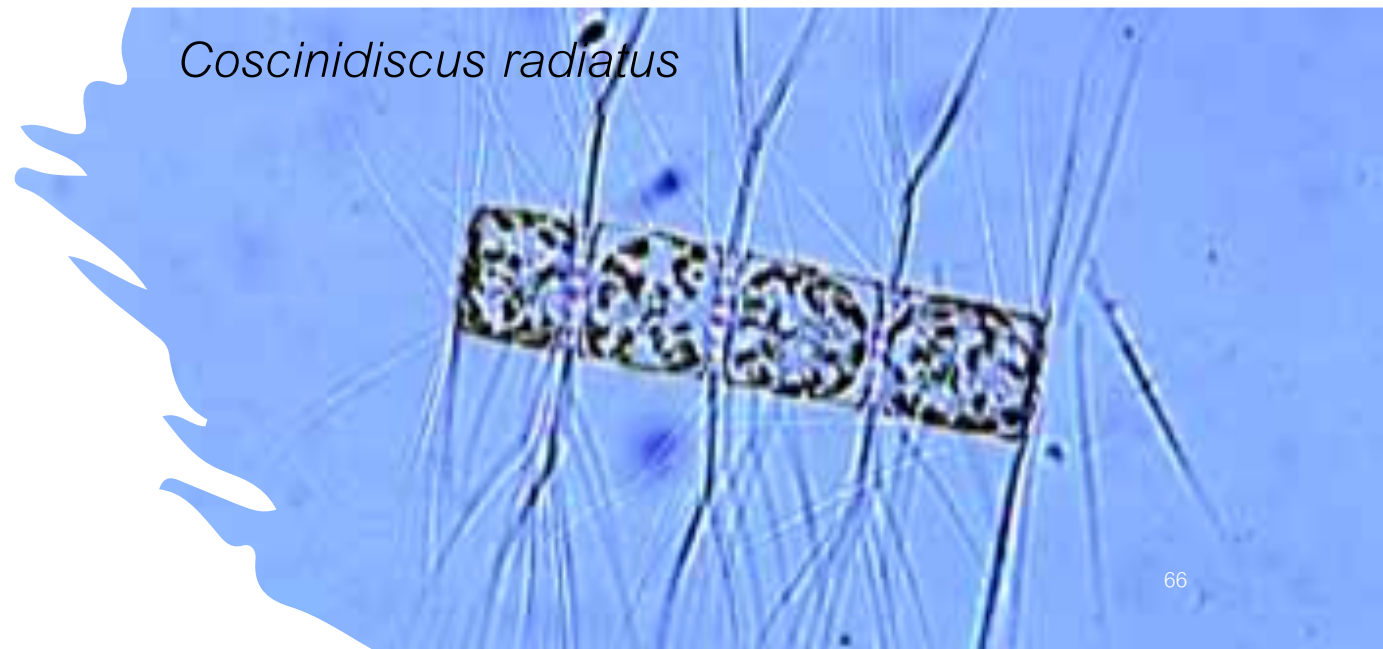


# Diatoms

Silica + high-nutrient conditions

# Diatom key traits

- A few hundred microns in size
- Have a hard silica frustule that contributes to their high sinking (contribute to high sinking flux)
- Dominate seasonal & local blooms because they grow quickly when opportunity strikes



# Diatoms can potentially manipulate their position in the water column

- They can move about by controlling how quickly they sink
- Being bulky, they would ordinarily sink quickly, but they may be able to *ballast* to keep themselves afloat
- Bulky glass shells may protect them from predators

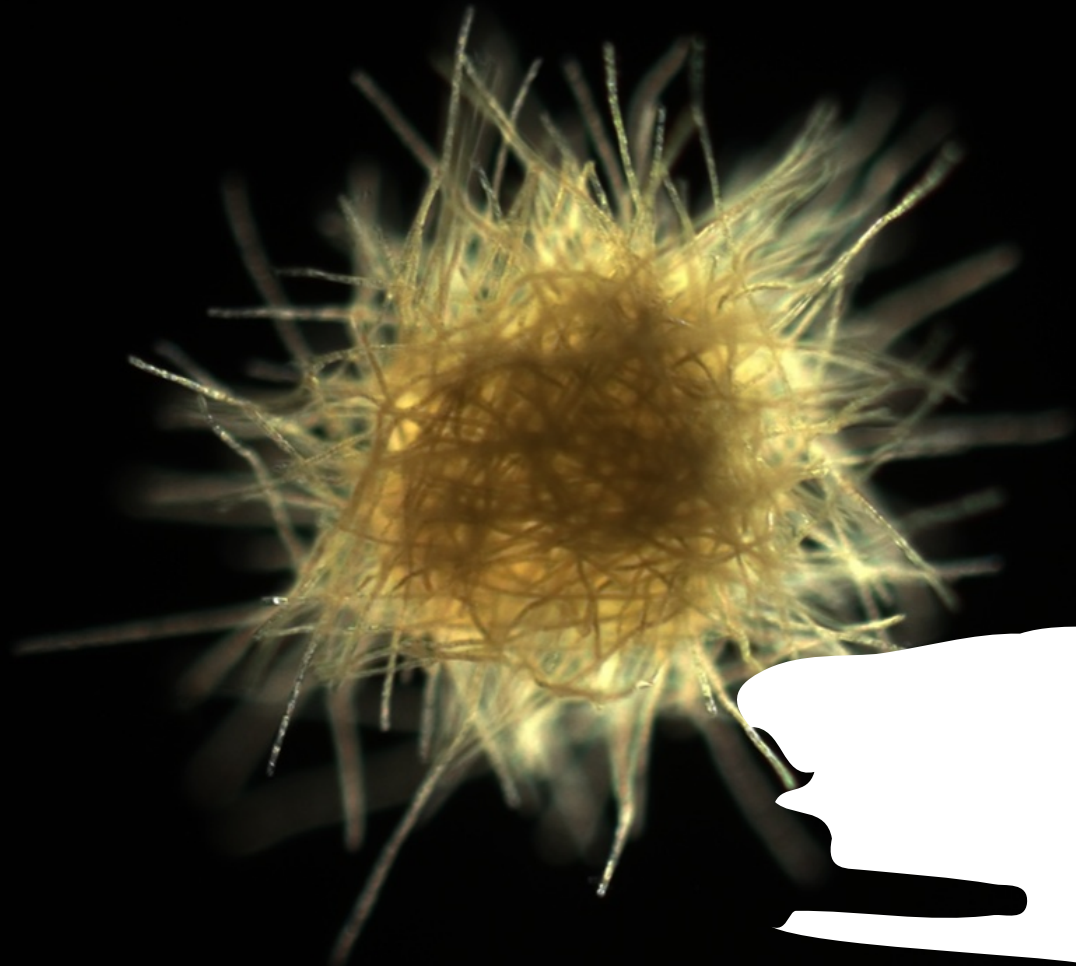


# The end of diatoms' life

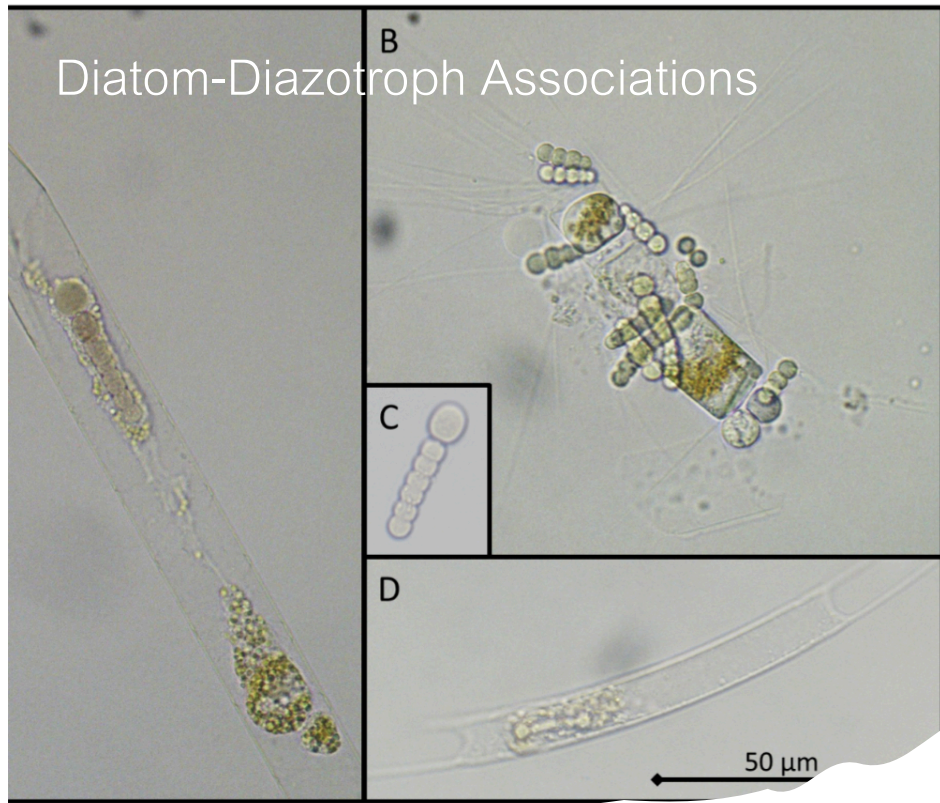
- As diatoms die, they sink and form diatomaceous earth material
- Chalky + lightweight
- Actually used to manufacture explosives
- You may have seen the material in the garden center for pest control when planting



Nitrogen fixers // diazotrophs



*Trichodesmium*  
Colony ~1mm



Diazotrophs take atmospheric nitrogen and make it usable

Diatoms & diazotrophs may have **symbiotic** relationships

# Ciliates



# Ciliates are important **heterotrophic** protists

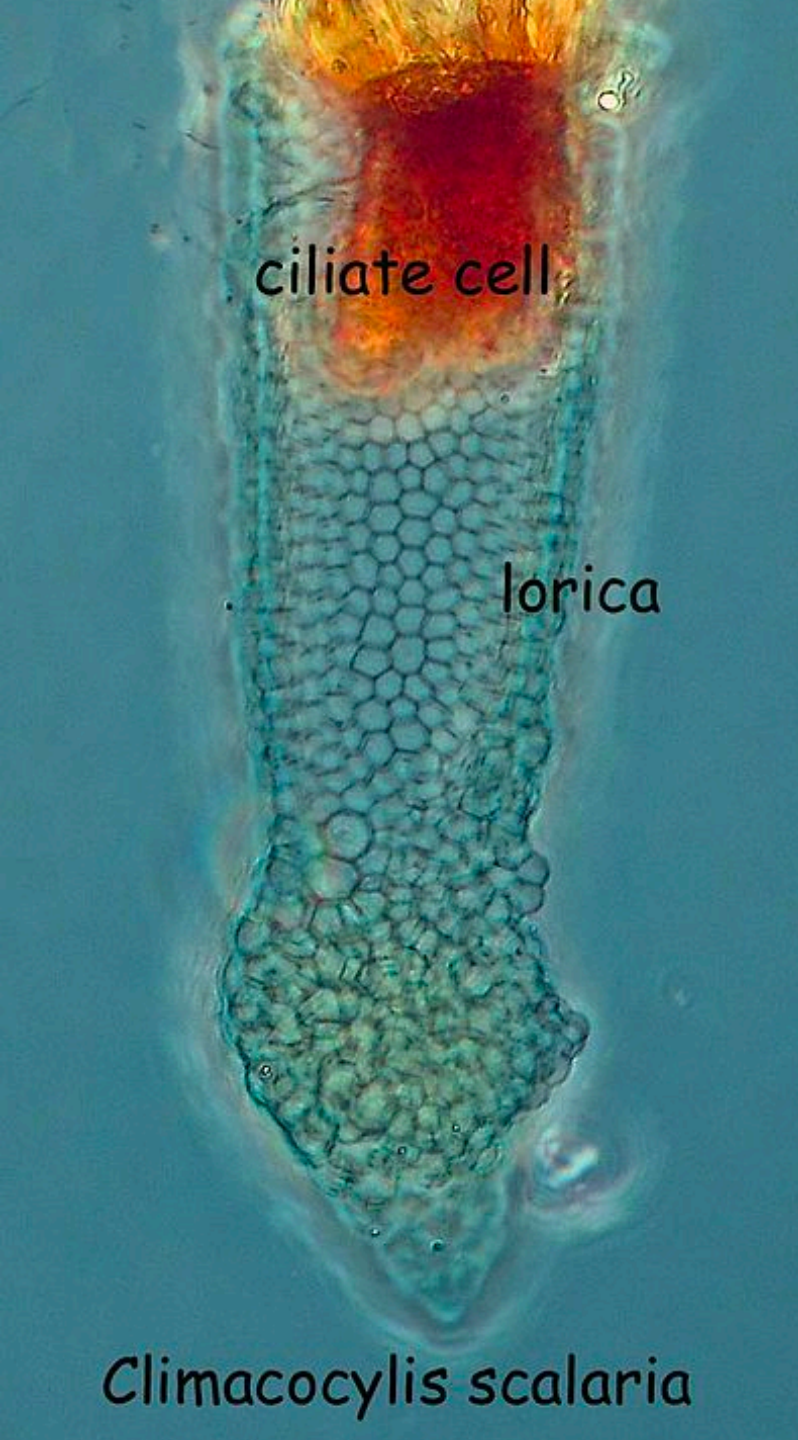
- Chow down on biomass of bacteria and smaller eukaryotic phytoplankton
- Form a key link in the microbial loop
- Also kleptoplastidic





## Oligotrichs

9/9/21



## Choreotrichs (including tintinnids)

- Lorica are the shells
- So, they're preserved like diatoms + coccolithophores
- Generally considered zooplankton/heterotrophs

Readings for next week!

# communications earth & environment




ARTICLE



<https://doi.org/10.1038/s43247-021-00201-y>

OPEN

## Photoacclimation by phytoplankton determines the distribution of global subsurface chlorophyll maxima in the ocean

Yoshio Masuda <sup>1,7</sup>✉, Yasuhiro Yamanaka<sup>1,8</sup>, Sherwood Lan Smith<sup>2,7</sup>, Takafumi Hirata<sup>3,8</sup>, Hideyuki Nakano<sup>4</sup>, Akira Oka <sup>5</sup> & Hiroshi Sumata <sup>6</sup>

- Reading 1: Masuda 2021 – more general paper

Readings for next week!



WILEY

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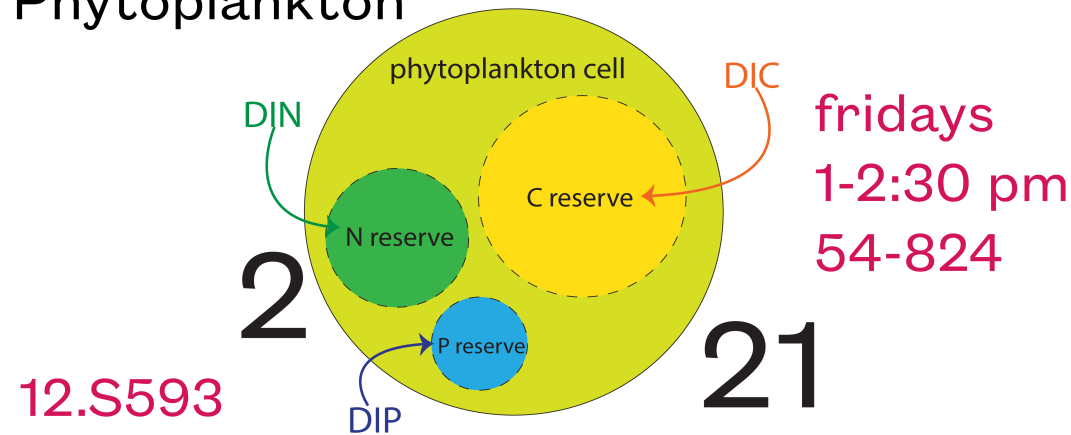
Light and Temperature Dependence of the Carbon to Chlorophyll a Ratio in Microalgae and Cyanobacteria: Implications for Physiology and Growth of Phytoplankton

Author(s): Richard J. Geider

Source: *The New Phytologist*, May, 1987, Vol. 106, No. 1 (May, 1987), pp. 1-34








- Reading 2: Geider 1987 – more model-specific

# Phytoplankton



## Physiology Seminar

this semester, we'll explore

1. out of thin air (light harvesting) 
2. dependencies (nutrients) 
3. on math, models, & traits 
4. setting a global stage 
5. I'll have it both ways (mixotrophy) 
6. teamwork makes the dream work (symbioses & competition) 
7. till death do us part (mortality) 
8. an aquatic pandemic? (viruses)
9. individual stoichiometry
10. an evolving process
11. biogeography & what's next?

from the lens of classic & modern literature

# Schedule of Readings

- Week 2
  - Geider 1987: Light and Temperature Dependence of the Carbon to Chlorophyll a Ratio in Microalgae and Cyanobacteria: Implications for Physiology and Growth of Phytoplankton
  - Masuda 2021: Photoacclimation by phytoplankton determines the distribution of global subsurface chlorophyll maxima in the ocean
- Week 3
  - Geider 2002: Redfield revisited: variability of C:N:P in marine microalgae and its biochemical basis
  - Morel 2008: The co-evolution of phytoplankton and trace element cycles in the oceans
- Week 4
  - Riley 1946: Factors Controlling Phytoplankton Populations on Georges Bank
  - Siegel 2002: The North Atlantic Spring Phytoplankton Bloom and Sverdrup's Critical Depth Hypothesis
- Week 5
  - Litchman 2015: Global biogeochemical impacts of phytoplankton: a trait-based perspective
  - Finkel 2009: Phytoplankton in a changing world: cell size and elemental stoichiometry
- Week 6
  - Stoecker 2017: Mixotrophy in the Marine Plankton
  - Caron 2016: Mixotrophy stirs up our understanding of marine food webs



# Schedule of Readings

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- Week 7
  - Burson 2018: Competition for nutrients and light: testing advances in resource competition with a natural phytoplankton community
  - Seymour 2017: Zooming in on the phycosphere: the ecological interface for phytoplankton–bacteria relationships
- Week 8
  - Hansen 1994: The size ration between planktonic predators and their prey
- Week 9
  - Menge 2009: Dangerous nutrients: Evolution of phytoplankton resource uptake subject to virus attack
  - Suttle 1990: Infection of phytoplankton by viruses and reduction of primary productivity
- Week 10
  - Inomura 2016: A quantitative analysis of the direct and indirect costs of nitrogen fixation: a model based on *Azotobacter vinelandii*
  - Klausmeier 2008: Phytoplankton stoichiometry



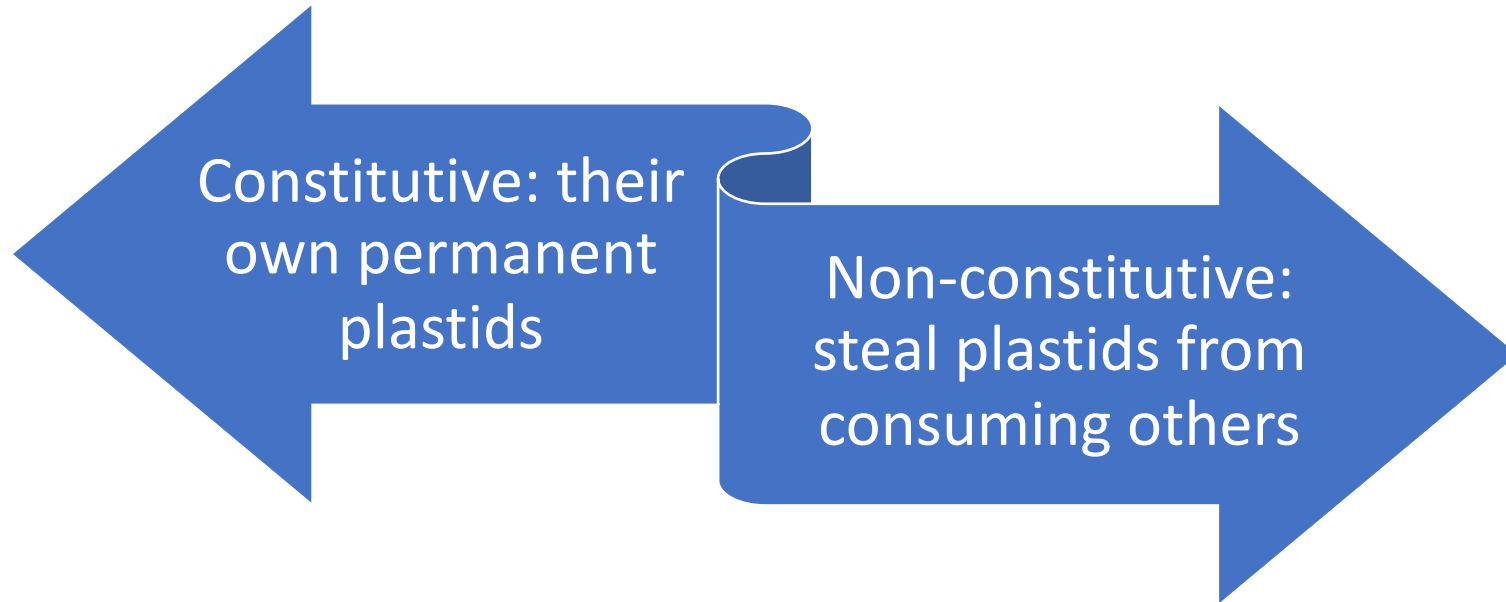


# Schedule of Readings

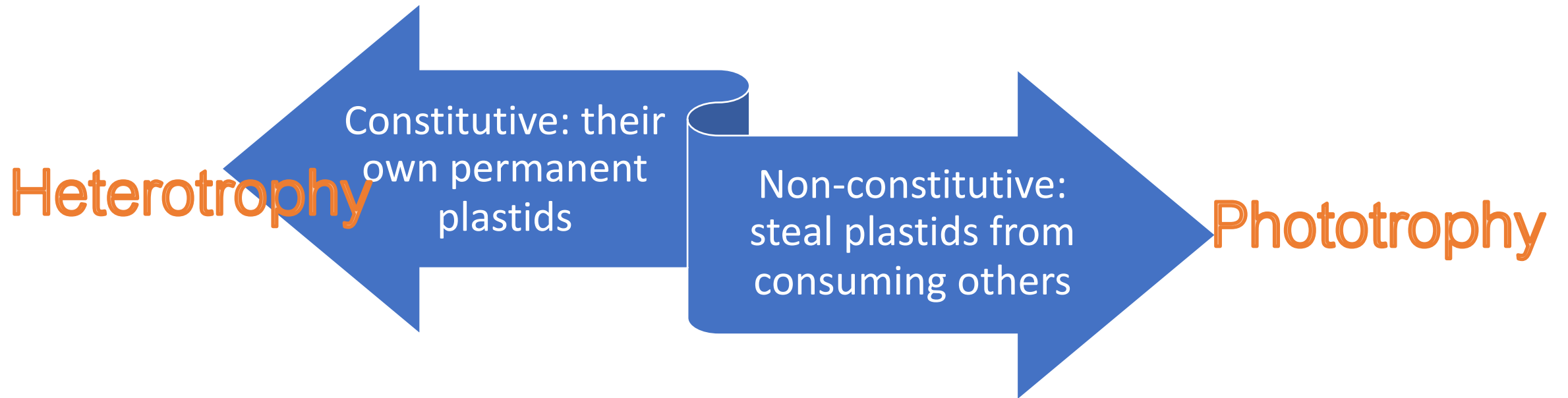
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- Week 11
  - Martiny 2020: Genomic adaptation of marine phytoplankton populations regulates phosphate uptake
- Week 12
  - Cermeno 2008: Resource levels, allometric scaling of population abundance, and marine phytoplankton diversity
  - Litchman 2008: Trait-based community ecology of phytoplankton

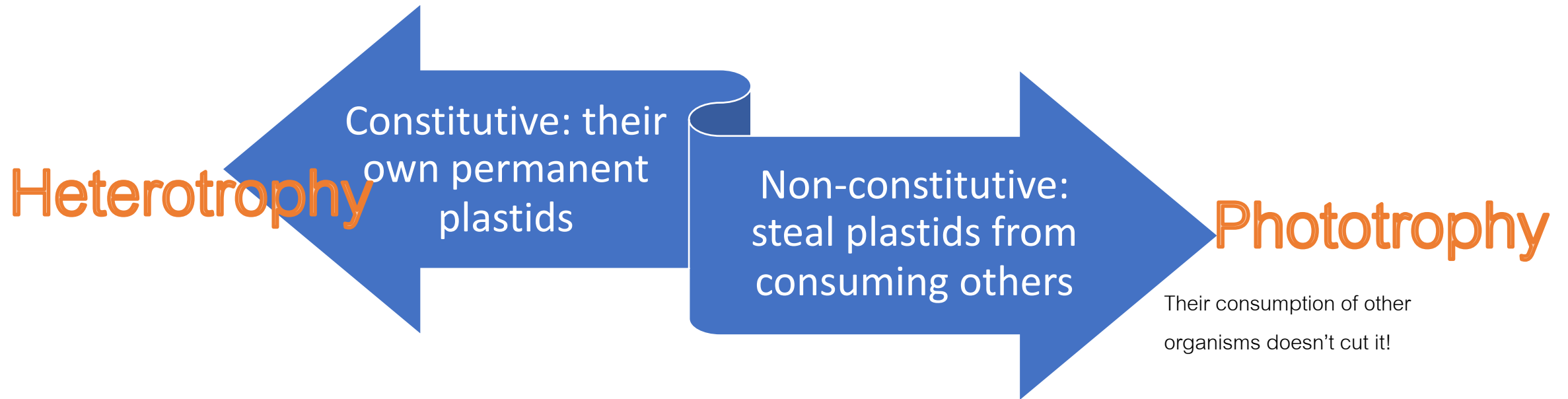
# Mixotrophy



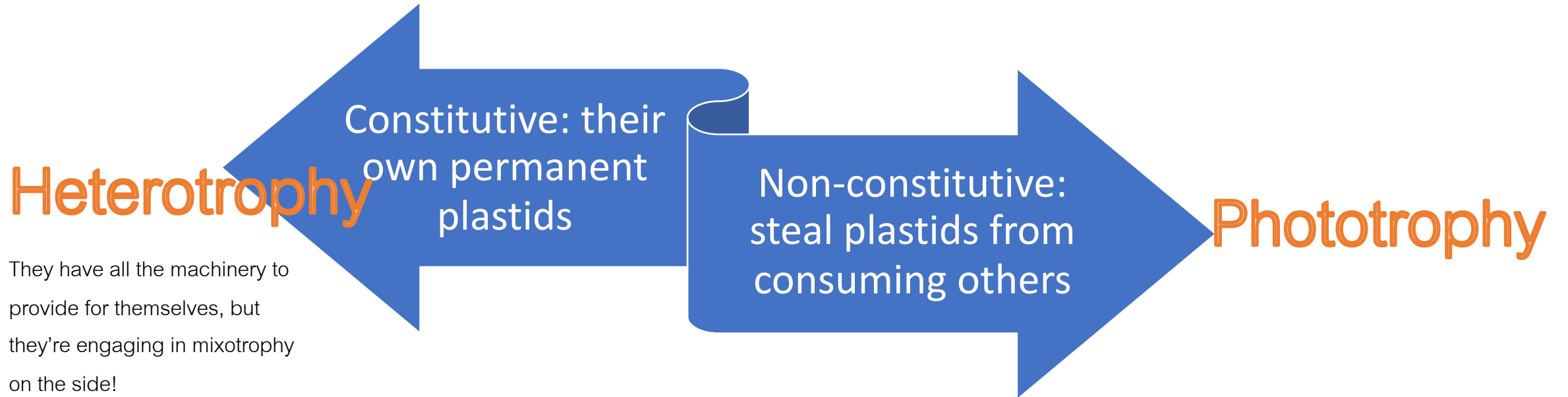
# Mixotrophy

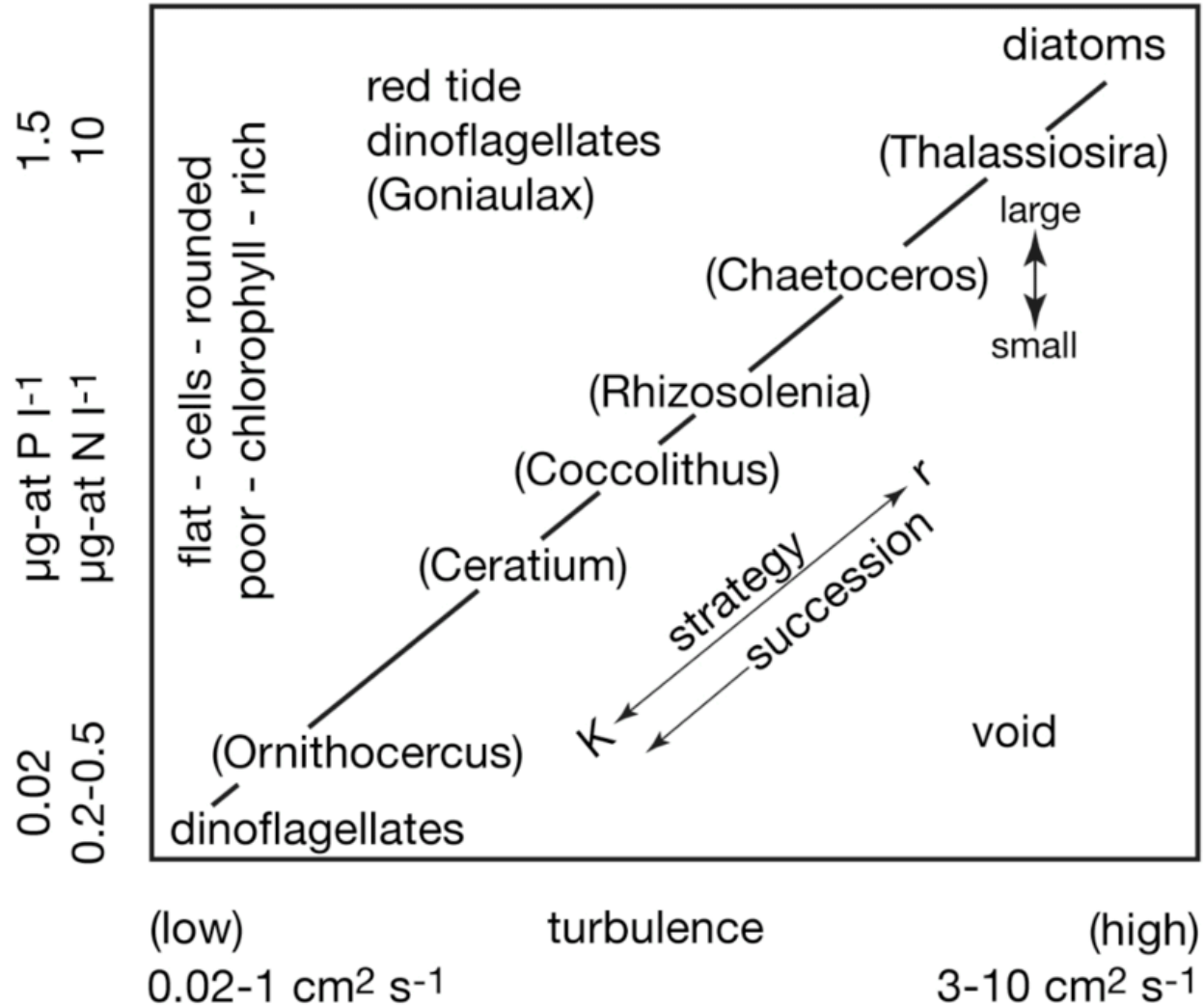


# Mixotrophy



# Mixotrophy





Margalef's Mandala

Plankton are in the Charles River, too!

